

Shishir P. S. Chundawat

Department of Chemical and Biochemical Engineering
98 Brett Road, Room C-150A
Rutgers, The State University of New Jersey
Piscataway, NJ 08854-8058
Office (Landline): +1-848-445-3678

Email: shishir.chundawat@rutgers.edu
Website: <http://chundawat.rutgers.edu>
Twitter: [@sps_chundawat](https://twitter.com/sps_chundawat)
Other Social Media: [LinkedIn](#), [Google Scholar](#), [ORCID](#), [Personal Zoom](#), [RUCore](#)

Education

Ph.D., Chemical Engineering, Michigan State University, December 2009
Dissertation Title: *Ultrastructural and physicochemical modifications within ammonia treated lignocellulosic cell walls and their influence on enzymatic digestibility*
Advisor - Prof. Bruce E. Dale

B. Tech. (Distinction), Chemical Technology, Institute of Chemical Technology, Mumbai, May 2004
Thesis Title: *Sucrose Esters - Manufacture, Analysis & Evaluation as Emulsifier-Detergent*
Advisor - Prof. D. N. Bhowmick

Professional Experience

2021-Present, Associate Professor (Tenured), Department of Chemical & Biochemical Engineering, Rutgers, The State University of New Jersey, New Brunswick

2015-2021, Assistant Professor (Tenure Track), Department of Chemical & Biochemical Engineering, Rutgers, The State University of New Jersey, New Brunswick

2012-2014, Assistant Staff Scientist, Great Lakes Bioenergy Research Center and Wisconsin Energy Institute, Department of Biochemistry, University of Wisconsin, Madison
Supervisor - Prof. Brian G. Fox

2011-2017, Adjunct/Research Assistant Professor (Non-Tenure Track), Department of Chemical Engineering & Materials Science, Michigan State University, East Lansing

2010-2011, Postdoctoral Research Associate, Department of Chemical Engineering & Materials Science, Michigan State University, East Lansing
Advisor - Prof. Bruce E. Dale

2008-Spring, Visiting Research Scholar, Verenum Cellulosic Biofuels (Enzymes Division), San Diego
Supervisor – Dr. Justin Stege

Research Interests and Keywords

Biochemical Engineering, Biomanufacturing, Biomass Process Engineering, Biopharmaceuticals, Bioseparations, Carbohydrate-Active enZymes (CAZymes), Cellulose & Carbohydrate Chemistry, Cellulosic Biofuels, Chemo-Enzymatic Synthesis, Glycans, Glycoconjugates, Glycoengineering, Glycobiology, Process Analytical Technology, Protein Adsorption, Protein Engineering, Single-Molecule Force Spectroscopy

Honors and Awards

2022: A. Walter Tyson Endowment Award, Sponsored by Rutgers University (SOE)
2020: Agilent Applications and Core Technology Faculty Award, Sponsored by Agilent Inc.
2019: National Science Foundation (NSF) Early Career Award, Sponsored by NSF
2018: A. Walter Tyson Assistant Professorship Award, Sponsored by Rutgers University (SOE)
2017: Undergraduate Teaching Excellence Award, Sponsored by Rutgers University (CBE)
2017: Outstanding Faculty of the Year, Sponsored by Rutgers University (CBE)
2016: Ralph E. Powe Junior Faculty Award, Sponsored by Oak Ridge Associated Universities (ORAU)
2010: Best 'Fundamental Science' Poster at 32nd Symposium of Biotechnology for Fuels and Chemicals
2009: Graduate School Fellowship Awardee, Sponsored by Michigan State University
2004: Government of Maharashtra (India) Merit Scholarship Holder from 2000-to-2004
2004: Mumbai University Bachelor of Technology Distinguished Rank Holder from 2000-to-2004
2003: ICT Alumni Association Best Undergraduate Award
2002: ICT Alumni Association Best Undergraduate Award
2001: J.G. Kane Memorial Trust Award

Peer-Reviewed Publications, Preprints, and Technical Reports

ORCID iD: <https://orcid.org/0000-0003-3677-6735>

Google Scholar: <https://scholar.google.com/citations?user=7Yf2K4IAAAAJ&hl=en>

Lead Corresponding Author on publications indicated as **Chundawat, S. P. S.***

Peer-Reviewed Papers (Black Font), Unrefereed Preprints or Technical Reports (Grey Font)

1. Seamann, A.; DeChellis, A.; **Chundawat, S. P. S.**; Khare S. ProteinMPNN informed supercharging – modifying the decoding mechanism of ProteinMPNN for sequence redesign. Manuscript submitted to ISCB Great Plains Bioinformatics Conference (in review). GitHub code available online (<https://github.com/Aseamann/mpnn-supercharging>).
2. DeChellis, A.; Trivedi, S.; Xie, L.; Khare, S.; **Chundawat, S. P. S.*** Supercharged Binding Modules Can Modulate Engineered Poly(Ethylene Terephthalate) Hydrolase Thermostability and Functional Persistence. bioRxiv. Cold Spring Harbor Laboratory May 27, 2026, p 2026.05.24.727315. <https://doi.org/10.64898/2026.05.24.727315>.

3. Rui, Y.; Zaoralova, M.; Dwyer, W. P.; Reyes, A. V.; Grismer, T. S.; Abel, N. B.; Jayachandran, D.; **Chundawat, S. P. S.**; Ott, T.; Kieber, J. J.; Dahlberg, P.; Xu, S.-L.; Dinneny, J. R. Cellulose Synthase Complexes and Remorins Mediate Stress Resilience Through Cell Wall-Plasma Membrane Attachments. *Cell*. **2026 (in press)**. <https://doi.org/10.1016/j.cell.2026.05.009>.
4. Nandi, S.; Stephens, T. G.; Garcia, R.; Sánchez-García, M.; Roberson, L. M.; Avalos, J. L.; **Chundawat, S. P. S.**; Bhattacharya, D. Rafts of Change: Microbial and Functional Dynamics in Simulated Sargassum Strandings. *Appl. Environ. Microbiol.* **2026**, 92 (4). <https://doi.org/10.1128/aem.02357-25>.
5. DeChellis, A.; Shimabukuro, S.; Trivedi, S.; Lubowski, R.; Nemmaru, B.; **Chundawat, S. P. S.*** There Is an “I” in Team: Individual Improvements in Supercharged Cellulase Cocktail Facilitates Cooperative Cellulose Degradation. *Biotechnol. Biofuels Bioprod.* **2026**, 19 (1), 14. <https://doi.org/10.1186/s13068-026-02740-y>.
6. DeChellis, A.; Shimabukuro, S.; Trivedi, S.; Lubowski, R.; Nemmaru, B.; **Chundawat, S. P. S.*** There Is an “I” in Team: Individual Improvements in Supercharged Cellulase Cocktail Facilitates Cooperative Cellulose Degradation. *bioRxiv*. Cold Spring Harbor Laboratory October 17, 2025, p 2025.10.16.682856. <https://doi.org/10.1101/2025.10.16.682856>.
7. Suaini, N. B.; Narvekar, A.; **Chundawat, S. P. S.*** Reaction Kinetics of Procainamide Dye Derivatization of N-Linked Glycans to Enable Robust Process Analytical Workflows for Glycoprotein-Based Biologics Manufacturing. *bioRxiv*. Cold Spring Harbor Laboratory October 15, 2025, p 2025.10.15.682640. <https://doi.org/10.1101/2025.10.15.682640>.
8. Rui, Y.; Zaoralova, M.; Dwyer, W. P.; Reyes, A. V.; Grismer, T. S.; Abel, N. B.; Jayachandran, D.; **Chundawat, S. P. S.**; Ott, T.; Kieber, J. J.; Dahlberg, P.; Xu, S.-L.; Dinneny, J. R. Cellulose Synthase Complexes and Remorins Mediate Stress Resilience Through Cell Wall-Plasma Membrane Attachments. *bioRxiv*. Cold Spring Harbor Laboratory August 2, 2025, p 2025.08.01.664786. <https://doi.org/10.1101/2025.08.01.664786>.
9. Gyorgypal, A.; Fratz-Berilla, E.; Kohnhorst, C.; Powers, D. N.; **Chundawat, S. P. S.*** Temporal Galactose-Manganese Feeding in Fed-Batch and Perfusion Bioreactors Modulates UDP-Galactose Pools for Enhanced MAb Glycosylation Homogeneity. *Biotechnol. Bioeng.* **2025**, 122, 1710–1723. <https://doi.org/https://doi.org/10.1002/bit.28999>.
10. Huh, H.; Jayachandran, D.; Sun, J.; Irfan, M.; Lam, E.*; **Chundawat, S. P. S.***; Lee, S.-H.* Time-Resolved Tracking of Cellulose Biosynthesis and Assembly during Cell Wall Regeneration in Live *Arabidopsis* Protoplasts. *Sci. Adv.* **2025**, 11 (12). <https://doi.org/10.1126/sciadv.ads6312>.
11. Dan, A.; Liu, B.; Patil, U.; Manuraj, B. N. M.; Gandhi, R.; Buchel, J.; **Chundawat, S. P. S.**; Guo, W.; Ramachandran, R*. Machine Learning Model-Based Design and Model Predictive Control of a Bioreactor for the Improved Production of Mammalian Cell-Based Bio-Therapeutics. *Control Eng. Pract.* **2025**, 156, 106198. <https://doi.org/10.1016/J.CONENGPRAC.2024.106198>.

12. Gyorgypal, A.; Chaturvedi, A.; Chopda, V.; Zhang, H.; **Chundawat, S. P. S.*** Evaluating the Impact of Media and Feed Combinations on CHO Cell Culture Performance and Monoclonal Antibody (Trastuzumab) Production. *Cytotechnology* **2025**, 77 (1), 40. <https://doi.org/10.1007/s10616-024-00690-7>.
13. Johnson, M. M.; DeChellis, A.; Nemmaru, B.; **Chundawat, S. P. S.**; Lang, M. J*. Thermobifida Fusca Cel6B Moves Bidirectionally While Processively Degrading Cellulose. *Biotechnol. Biofuels Bioprod.* **2024**, 17 (1), 140. <https://doi.org/10.1186/s13068-024-02588-0>.
14. Jayachandran, D.; Parvate, A. D.; Brookreson, J. T.; Evans, J. E.*; **Chundawat, S. P. S.*** Cell-Free Expression and Biochemical Characterization of Polysaccharide-Synthesizing Glycosyltransferases. *Biochem. Eng. J.* **2024**, 212, 109507. <https://doi.org/10.1016/J.BEJ.2024.109507>.
15. Huh, H.; Jayachandran, D.; Sun, J.; Irfan, M.; Lam, E.*; **Chundawat, S. P. S.***; Lee, S.-H*. Time-Resolved Tracking of Cellulose Biosynthesis and Microfibril Network Assembly during Cell Wall Regeneration in Live Arabidopsis Protoplasts. *bioRxiv* **2024**, 2024.07.29.605709. <https://doi.org/10.1101/2024.07.29.605709>.
16. Nemmaru, B., Douglass, J., Yarbrough, J. M., DeChellis, A., Shankar, S., Thokkadam, A., Wang, A. & **Chundawat, S. P. S.*** Supercharged cellulases show superior thermal stability and enhanced activity towards pretreated biomass and cellulose. *Frontiers in Energy Research.* **2024**, 12, 1372916. <https://www.frontiersin.org/articles/10.3389/fenrg.2024.1372916/abstract>. **Invited article for special journal research topic issue titled “10 Years of Frontiers in Energy Research”.**
17. Jayachandran, D.; Parvate, A. D.; Brookreson, J. T.; Evans, J. E.*; **Chundawat, S. P. S.*** Cell-Free Expression and Biochemical Characterization of Polysaccharide-Synthesizing Glycosyltransferases. *bioRxiv* **2024**, 2024.02.12.580006. <https://doi.org/10.1101/2024.02.12.580006>.
18. DeChellis, A.; Nemmaru, B.; Sammond, D.; Douglass, J.; Patil, N.; Reste, O.; **Chundawat, S. P. S.*** Supercharging Carbohydrate Binding Module Alone Enhances Endocellulase Thermostability, Binding, and Activity on Cellulosic Biomass. *ACS Sustain. Chem. Eng.* **2024**, 12, 9, 3500-3516. <https://doi.org/10.1021/acssuschemeng.3c06266>.
19. Ramdin, K.; Hackl, M.; **Chundawat, S. P. S.*** Visualization of Tethered Particle Motion with a Multidimensional Simulation. *Biophys.* **2024**. <https://doi.org/10.35459/tbp.2022.000238>.
20. Jayachandran, D.; Banerjee, S.; **Chundawat, S. P. S.*** Plant Cellulose Synthase Membrane Protein Isolation Directly from Pichia Pastoris Protoplasts, Liposome Reconstitution, and Its Enzymatic Characterization. *Protein Expr. Purif.* **2023**, 210, 106309. <https://doi.org/10.1016/J.PEP.2023.106309>.
21. Nemmaru, B.; DeChellis, A.; Patil, N.; **Chundawat, S. P. S.*** CAZyme Characterization and Engineering for Biofuels Applications. In *Handbook of Biorefinery Research and Technology*;

Springer Netherlands: Dordrecht, 2023; pp 1–34. https://doi.org/10.1007/978-94-007-6724-9_32-1. **Invited Book Review Chapter.**

22. Jayachandran, D.; Smith, P.; Irfan, M.; Sun, J.; Yarborough, J. M.; Bomble, Y. J.; Lam, E.; **Chundawat, S. P. S.*** Engineering and Characterization of Carbohydrate-binding Modules for Imaging Cellulose Fibrils Biosynthesis in Plant Protoplasts. *Biotechnol. Bioeng.* **2023**, *120* (8), 2253–2268. <https://doi.org/10.1002/bit.28484>.
23. Lee, S.-H.*; **Chundawat, S.***; Lam, E.; Lang, M.; Muchero, W.; Vankatesh Pingali, S. *In Planta Single-Molecule Imaging and Holographic Force Spectroscopy to Study Real-Time, Multimodal Turnover Dynamics of Polysaccharides and Associated Carbohydrate Metabolites*; Argonne, IL (United States), **2023**. Technical Report for USDOE Office of Science (SC), Biological and Environmental Research (BER). <https://doi.org/10.2172/1960742>.
24. Roy, S.; **Chundawat, S. P. S.** Ionic Liquid–Based Pretreatment of Lignocellulosic Biomass for Bioconversion: A Critical Review. *BioEnergy Res.* **2023**, *16*, 263–278. <https://doi.org/10.1007/s12155-022-10425-1>.
25. Hackl, M.; Power, Z.; **Chundawat, S.P.S.*** Oriented Display of Cello-Oligosaccharides for Pull-down Binding Assays to Distinguish Binding Preferences of Glycan Binding Proteins. *Carbohydr. Res.* **2023**, *534*, 108943. <https://doi.org/10.1016/J.CARRES.2023.108943>.
26. Hackl, M.; Power, Z.; **Chundawat, S.P.S.*** Oriented Display of Cello-Oligosaccharides for Pull-down Binding Assays to Distinguish Binding Preferences of Glycan Binding Proteins. *bioRxiv* **2023**, 2023.01.26.525732. <https://doi.org/10.1101/2023.01.26.525732>.
27. Tiwold, E. K.; Gyorgypal, A.; **Chundawat, S. P. S.*** Recent Advances in Biologic Therapeutic N-Glycan Preparation Techniques and Analytical Methods for Facilitating Biomanufacturing Automation. *J. Pharm. Sci.* **2023**, *112* (6), 1485–1491. <https://doi.org/10.1016/j.xphs.2023.01.012>.
28. Jayachandran, D.; Smith, P.; Irfan, M.; Sun, J.; Yarborough, J. M.; Bomble, Y. J.; Lam, E.; **Chundawat, S. P. S.*** Engineering and Characterization of Carbohydrate-Binding Modules to Enable Real-Time Imaging of Cellulose Fibrils Biosynthesis in Plant Protoplasts. *bioRxiv* **2023**, 2023.01.02.522519. <https://doi.org/10.1101/2023.01.02.522519>.
29. Ganesan, G.; Hackl, M.; **Chundawat, S. P. S.*** Characterizing the Cellulose Binding Interactions of Type-A Carbohydrate-Binding Modules Using Acoustic Force Spectroscopy. *bioRxiv* **2023**, 2023.10.18.563009. <https://doi.org/10.1101/2023.10.18.563009>.
30. DeChellis, A.; Nemmaru, B.; Sammond, D.; Douglas, J.; Patil, N.; Reste, O.; **Chundawat, S. P. S.*** Supercharging Carbohydrate-Binding Module Alone Enhances Endocellulase Thermostability, Binding, and Activity on Cellulosic Biomass. *bioRxiv* **2023**, 2023.09.09.557007. <https://doi.org/10.1101/2023.09.09.557007>.

31. Gyorgypal, A.; Potter, O. G.; Chaturvedi, A.; Powers, D. N.; **Chundawat, S. P. S.*** Automated Instant Labeling Chemistry Workflow for Real-Time Monitoring of Monoclonal Antibody N-Glycosylation. *React. Chem. Eng.* **2023**, *8*, 2423–2434. <https://doi.org/10.1039/D2RE00568A>. **Invited Article to Special Issue on Biocatalysis and Bioprocessing in RSC Reaction Chemistry and Engineering.**
32. Gyorgypal, A.; Fratz-Berilla, E.; Kohnhorst, C.; Powers, D. N.; **Chundawat, S. P. S.*** Temporal Effects of Galactose and Manganese Supplementation on Monoclonal Antibody N-Linked Glycosylation in Fed-Batch and Perfusion Bioreactor Operation. *bioRxiv* **2023**, 2023.04.15.535602. <https://doi.org/10.1101/2023.04.15.535602>.
33. Hackl, M.; Jayachandran, D.; Ramdin, K.; Zhong, T.; **Chundawat, S. P. S.*** Site-Specific Effector Protein Functionalization to Create Bead-Based Avidity Model Systems. *bioRxiv* **2023**, 2023.03.27.534459. <https://doi.org/10.1101/2023.03.27.534459>.
34. Kumar, M.; Bandi, C. K.; **Chundawat, S.P.S.*** High-Throughput Screening of Glycosynthases Using Azido Sugars for Oligosaccharides Synthesis. *Methods Enzymol.* **2023**, *682*, 211–245. <https://doi.org/10.1016/BS.MIE.2022.12.002>. **Invited Article in Integrated Methods in Protein Biochemistry: Part C Series of Methods in Enzymology Edited by Arun K. Shukla.**
35. Gyorgypal, A.; Potter, O.; Chaturvedi, A.; Powers, D. N.; **Chundawat, S. P. S.*** Automated Workflow for Instant Labeling and Real-Time Monitoring of Monoclonal Antibody N-Glycosylation. *bioRxiv* **2022**, 2022.12.22.521623. <https://doi.org/10.1101/2022.12.22.521623>.
36. Guranovic, I.; Kumar, M.; Bandi, C. K.; **Chundawat, S. P. S.*** Machine Learning Assisted Ligand Binding Energy Prediction for in Silico Generated Glycosyl Hydrolase Enzyme Combinatorial Mutant Library. *bioRxiv* **2022**, 2022.11.29.518414. <https://doi.org/10.1101/2022.11.29.518414>.
37. Hackl, M.; Contrada, E. V.; Ash, J. E.; Kulkarni, A.; Yoon, J.; Cho, H.-Y.; Lee, K.-B.; Yarbrough, J. M.; López, C. A.; Gnanakaran, S.; **Chundawat, S. P. S.*** Acoustic Force Spectroscopy Reveals Subtle Differences in Cellulose Unbinding Behavior of Carbohydrate-Binding Modules. *Proc. Natl. Acad. Sci.* **2022**, *119* (42), e2117467119. <https://doi.org/10.1073/pnas.2117467119>.
38. Ramdin, K. A.; Hackl, M.; **Chundawat, S. P. S.*** Multivariable Graphical User Interface for Simulation of Tethered Particle Motion. *bioRxiv* **2022**, 2022.08.31.506066. <https://doi.org/10.1101/2022.08.31.506066>.
39. Chopda, V., Gyorgypal, A., Yang, O., Singh, R., Ramachandran, R., Zhang, H., Tsilomelekis, G., **Chundawat, S. P. S.***, and Ierapetritou, M. G. Recent Advances in Integrated Process Analytical Techniques, Modeling, and Control Strategies to Enable Continuous Biomanufacturing of Monoclonal Antibodies. *Journal of Chemical Technology & Biotechnology*, **2022**, *97* (9), 2317–2335. <https://doi.org/10.1002/jctb.6765>.
40. Tiwold, E. K.; Gyorgypal, A.; **Chundawat, S. P. S.*** Recent Advances in Biologic Therapeutic N-Glycan Preparation Techniques and Analytical Methods for Facilitating Biomanufacturing Automation.

ChemRxiv (Cambridge Cambridge Open Engag). **2022**. <https://doi.org/10.26434/chemrxiv-2022-sdq2g>.

41. Gyorgypal, A.; **Chundawat, S. P. S.***. Integrated Process Analytical Platform for Automated Monitoring of Monoclonal Antibody N-Linked Glycosylation. *Anal. Chem.* **2022**, 94 (19), 6986–6995. <https://doi.org/10.1021/acs.analchem.1c05396>. **Journal Cover Article**.
42. Hackl, M., Contrada, E. V., Ash, J. E., Kulkarni, A., Yoon, J., Cho, H.-Y., Lee, K.-B., Yarbrough, J. M. & **Chundawat, S. P. S.***, "Acoustic Force Spectroscopy Reveals Subtle Differences in Cellulose Unbinding Behavior of Carbohydrate-Binding Modules," *bioRxiv* **2021**, 2021.09.20.461102. <https://doi.org/10.1101/2021.09.20.461102>
43. Nemmaru, B., Douglass, J., Yarbrough, J. M., Chellis, A. De, Shankar, S., Thokkadam, A., Wang, A. & **Chundawat, S. P. S.***, "Supercharged Cellulases Show Reduced Non-Productive Binding, But Enhanced Activity, on Pretreated Lignocellulosic Biomass," *bioRxiv* **2021**, 2021.10.17.46468. <https://doi.org/10.1101/2021.10.17.464688>
44. Gyorgypal, A. & **Chundawat, S. P. S.***, "An Integrated Process Analytical Platform for Automated Monitoring of Monoclonal Antibody N-linked Glycosylation," *bioRxiv* **2021**, 2021.11.14.468439. <https://doi.org/10.1101/2021.11.14.468439>
45. Agrawal, A., Bandi, C. K., Burgin, T., Woo, Y., Mayes, H. B., and **Chundawat, S. P. S.*** "Click-Chemistry-Based Free Azide versus Azido Sugar Detection Enables Rapid In Vivo Screening of Glycosynthase Activity." *ACS Chemical Biology*, **2021**, p. acschembio.1c00585. <https://doi.org/10.1021/acschembio.1c00585>.
46. Bandi, C. K., Skalenko, K. S., Agrawal, A., Sivaneri, N., Thiry, M., and **Chundawat, S. P. S.*** Engineered Regulon to Enable Autonomous Azide Ion Biosensing, Recombinant Protein Production, and in Vivo Glycoengineering. *ACS Synthetic Biology*, Vol. 10, No. 4, **2021**, pp. 682–689. <https://doi.org/10.1021/acssynbio.0c00449>.
47. Nemmaru, B., Ramirez, N., Farino, C. J., Yarbrough, J. M., Kravchenko, N., and **Chundawat, S. P. S.*** Reduced Type-A Carbohydrate-binding Module Interactions to Cellulose I Leads to Improved Endocellulase Activity. *Biotechnology and Bioengineering*, Vol. 118, No. 3, **2020**, p. bit.27637. <https://doi.org/10.1101/2020.07.02.183293>.
48. **Chundawat, S. P. S.***, Nemmaru, B., Hackl, M., Brady, S. K., Hilton, M. A., Johnson, M. M., Chang, S., Lang, M. J., Huh, H., Lee, S.-H., Yarbrough, J. M., López, C. A., and Gnanakaran, S. Molecular Origins of Reduced Activity and Binding Commitment of Processive Cellulases and Associated Carbohydrate-Binding Proteins to Cellulose III. *The Journal of Biological Chemistry*, Vol. 296, **2021**, p. 100431. <https://doi.org/10.1016/j.jbc.2021.100431>.

49. Bandi, C. K., Agrawal, A., and **Chundawat, S. P. S.*** Carbohydrate-Active EnZyme (CAZyme) Enabled Glycoengineering for a Sweeter Future. *Current Opinion in Biotechnology*, **2020**, Volume 66, 283–291. <https://doi.org/10.1016/j.copbio.2020.09.006>. **Invited review article.**
50. Bandi, C. K., Goncalves, A., Pingali, S. V., and **Chundawat, S. P. S.*** Carbohydrate-binding Domains Facilitate Efficient Oligosaccharides Synthesis by Enhancing Mutant Catalytic Domain Transglycosylation Activity. *Biotechnology and Bioengineering*, Vol. 117, **2020**, pp. 2944– 2956. <https://doi.org/10.1002/bit.27473>.
51. Bandi, C. K.; Skalenko, K. S.; Agrawal, A.; Sivaneri, N.; Thiry, M.; **Chundawat, S. P. S.*** Synthetic Promoter Based Azide Biosensor Toolkit to Advance Chemical-Biology. *bioRxiv*, **2020**, 2020.07.08.193060. <https://doi.org/10.1101/2020.07.08.193060>.
52. **Chundawat, S. P. S.*** Comparison of Analytical Methods for Rapid and Reliable Quantification of Plant-Based Carbohydrates for the Quintessential Bioenergy Educator. *bioRxiv*, **2020**, 2020.05.21.106468. <https://doi.org/10.1101/2020.05.21.106468>.
53. **Chundawat, S. P. S.***, Nemmaru, B., Hackl, M., Brady, S. K., Hilton, M. A., Johnson, M. M., Chang, S., Lang, M. J., Huh, H., Lee, S.-H., Yarbrough, J. M., López, C. A., and Gnanakaran, S., “Multiscale Characterization of Complex Binding Interactions of Cellulolytic Enzymes Highlights Limitations of Classical Approaches,” *bioRxiv*, **2020**, p. 2020.05.08.084152. <https://www.biorxiv.org/content/10.1101/2020.05.08.084152v1.full>.
54. Nemmaru, B.; Ramirez, N.; Farino, C. J.; Yarbrough, J. M.; Kravchenko, N.; **Chundawat, S. P. S.*** Reduced Type-A Carbohydrate-Binding Module Interactions to Cellulose Leads to Improved Endocellulase Activity. *bioRxiv*, **2020**, 2020.07.02.183293. <https://doi.org/10.1101/2020.07.02.183293>.
55. **Chundawat, S. P. S.***, Pal, R., Zhao, C., Campbell, T., Teymouri, F., Videto, J., Nielson, C., Wiefelrich, B., Sousa, L., Dale, B. E., Balan, V., Chipkar, S., Aguado, J., Burke, E., and Ong, R. G., “Ammonia Fiber Expansion (AFEX) Pretreatment of Lignocellulosic Biomass,” *J. Vis. Exp*, **2020**, p. 57488. <https://www.jove.com/video/57488>.
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57. Liu, Y., Nemmaru, B., and **Chundawat, S. P. S.***, “Thermobifida fusca Cellulases Exhibit Increased Endo–Exo Synergistic Activity, but Lower Exocellulase Activity, on Cellulose-III,” *ACS Sustainable Chemistry & Engineering*, **2020**, p. acssuschemeng.9b06792. <https://pubs.acs.org/doi/10.1021/acssuschemeng.9b06792>.

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59. **Chundawat, S. P. S.***, Sousa, L. daCosta, Roy, S., Yang, Z., Gupta, S., Pal, R., Zhao, C., Liu, S.-H., Petridis, L., O'Neill, H., and Pingali, S. V., "Ammonia-salt solvent promotes cellulosic biomass deconstruction under ambient pretreatment conditions to enable rapid soluble sugar production at ultra-low enzyme loadings," *Green Chemistry*, vol. 22, **2020**, pp. 204–218.
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Patents

2026: Electrostatic tuning of CBM fusions modulates thermostability and functional persistence of PET hydrolases (U.S. Provisional Application pending; Rutgers Notice of Invention DISC-2026-847 filed on May 15, 2026). Inventors: **Shishir Chundawat**, Antonio DeChellis.

2026: Saltwater pretreatment of *Sargassum* brown algae and halophilic conditions for enzymatic deconstruction (U.S. Provisional Application pending; Rutgers Notice of Invention DISC-2026-824 filed on April 23, 2026). Inventors: **Shishir Chundawat**, Rebecca Garcia, Mitun Aghipothu Umashankar.

2025: Glycosylated monoclonal antibody critical quality attributes determination using fully automated UHPLC sample preparation and analytical workflows (U.S. Provisional Application pending; Rutgers Notice of Invention DISC-2025-299 filed on June 4, 2025). Inventors: **Shishir Chundawat**, Aditya Narvekar.

2023: Compositions comprising glycoside hydrolase enzyme variants which exhibit improved cellulose hydrolysis and methods of use thereof (U.S. Provisional Application 63/491,331 filed on March 21, 2023). Inventors: **Shishir Chundawat**, Bhargava Nemmaru, Antonio DeChellis, Deanne Sammond.

2022: Automated real-time, on-line N-glycosylation monitoring methods and systems thereof (US Patent Application No. US20240110925A1 filed Sept 30, 2022). Inventors: **Shishir Chundawat**, Aron Gyorgypal. <https://patents.google.com/patent/US20240110925A1>

2020: Biosensors for Selectively Identifying Azide Ions (US Patent Application No. US20230279464A1 filed June 30, 2020). Inventors: **Shishir Chundawat**, Chandra Kanth Bandi, Kyle Skalenko. <https://patents.google.com/patent/US20230279464A1/en>

2019: High-Throughput Cell-Based Screening Methodology for Evaluating Carbohydrate-Active Enzymes (US Patent Application No. US20210024974A1, filed July 22, 2019). Inventors: **Shishir Chundawat**, Chandra Kanth Bandi, Ayushi Agrawal.

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2019: Engineered Carbohydrate-Active Enzymes for Glycan Polymers Synthesis (US Patent 11,248,217). Inventors: **Shishir Chundawat**, Chandra Kanth Bandi.

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2017: Methods for producing extracted and digested products from pretreated lignocellulosic biomass (US Patent 9,650,657 B2). Inventors: **Shishir Chundawat**, Leonardo Da Costa Sousa, Albert M. Cheh, Venkatesh Balan, Bruce Dale. <https://patents.google.com/patent/US9650657B2>.

2017: Methods for pretreating biomass (US Patent 9,644,222). Inventors: Venkatesh Balan, Bruce Dale, **Shishir Chundawat**, Leonardo Sousa. <https://patents.google.com/patent/US9644222B2>.

2015: Extraction of solubles from plant biomass for use as microbial growth stimulant and methods related thereto (US Patent 9,206,446 B2). Inventors: Ming Woei Lau, Bruce Dale, Venkatesh Balan, **Shishir Chundawat**. <https://patents.google.com/patent/US9206446B2>.

2013: Fractionated extractive products from plant biomass and methods of making and using same (US Patent Application 20130244293 A1). Inventors: Venkatesh Balan, Albert Cheh, **Shishir Chundawat**, Bruce E. Dale, Leonardo Sousa. <https://patents.google.com/patent/US20130244293A1>.

2013: Process for producing sugars and ethanol using corn stillage (US Patent 8,367,378 B2). Inventors: Venkatesh Balan, **Shishir Chundawat**, Leonardo Sousa, Bruce Dale.

<https://patents.google.com/patent/US8367378B2>.

2009: Process for making fuels and chemicals from AFEX treated whole grain or whole plants (US Patent Application 20090053771 A1). Inventors: Bruce Dale, Venkatesh Balan, **Shishir Chundawat**, Bryan Bals. <https://patents.google.com/patent/US20090053771A1>.

2009: Process for enzymatically converting a plant biomass (US Patent Application 20090042259 A1). Inventors: Bruce E. Dale, Farzaneh Teymouri, **Shishir Chundawat**, Venkatesh Balan.

<https://patents.google.com/patent/US20090042259A1>.

2007: Composite materials from corncob granules and process for preparation (US Patent Application 20070287795 A1). Inventors: Masud Huda, Venkatesh Balan, Lawrence Drzal, Bruce Dale, **Shishir Chundawat**, Manjusri Misra. <https://patents.google.com/patent/US20070287795A1>.

2007: Process for conversion of mushroom lignocellulosic waste to useful byproducts (US Patent Application 20070227063 A1). Inventors: Bruce E. Dale, Venkatesh Balan, **Shishir Chundawat**.

<https://patents.google.com/patent/US20070227063A1>.

Research Funding

Externally Funded Research and User Facility Grants

Current Projects

Project Title: *Targeted high-throughput design of biocatalysts for degrading selected polymers*

Award Number: TBD (DOE-JGI Community Science Program)

Grant Amount: Upto 500 kbp total DNA synthesis

Start Date: Sept 2024, Tentative End Date: Dec 2028

Investigators: Sagar Khare (Rutgers Lead PI), **Shishir Chundawat** (Co-PI)

Agency: Department of Energy (Joint Genome Institute Annual DNA Synthesis Award)

Role: Co-PI

Project Title: *Commercial Deployment of Integrated PAT Technologies to Enable Continuous Upstream Production of Glycosylated Monoclonal Antibodies*

Award Number: PC7.1-146

Grant Amount: \$1,937,493 (including cost share for industry personnel time and donated equipment)

Start Date: Dec 2024, Tentative End Date: Aug 2026

Investigators: **Shishir Chundawat** (Rutgers, Lead PI), Randall Robinson (Agilent, Co-I), Sean Gilliam (Endress+Hauser, Co-I), Shilpa Gadgil (Enzene Inc, Co-I), Kevin Lee (Millipore-Sigma, Co-I), Guogang Dong (Pfizer, co-I)

Agency: National Institute for Innovation in Manufacturing Biopharmaceuticals (NIIMBL)

Role: PI

Project Title: *Bioconversion of low-value dairy sugars to high-value prebiotics*

Award Number: 4061-0

Grant Amount: \$100,000

Start Date: May 2024, Tentative End Date: Aug 2026

Investigators/Institutions: **Shishir Chundawat** (PI, Rutgers)

Agency: National Dairy Council

Role: PI

Project Title: *Sargassum seaweed as a future renewable feedstock for sustainable biomanufacturing*

Award Number: Sargassum BioRefinery (SaBRe) Center Award (G-24-66434)

Grant Amount: \$10,433,481

Start Date: April 2024, End Date: June 2029

Principal Investigators/Institutions: Jose Avalos (PI, Princeton), **Shishir Chundawat** (Co-PI, Rutgers), Loretta Roberson (co-PI, Marine Biological Laboratory)

Collaborators/Institutions: Sagar Khare (Rutgers), Debashish Bhattacharya (Rutgers), Yalin Li (Rutgers), Liz Diaz (University of Puerto Rico), Alissa Park (UCLA), Aaron Moment (UCLA), Hauke Kite-Powell (Woods Hole Oceanographic Institution), Daniel Robledo (Cinvestav-Merida), Yasuo Yoshikuni (LBNL), Carbonwave.

Agency: Schmidt Sciences (Virtual Institute on Feedstocks of the Future)

Role: SaBRe Center Co-PI, Rutgers Lead PI

Project Title: *CAREER-Force Spectroscopy Enabled Multivalent Glycan-binding Protein Engineering*

Award Number: 1846797

Grant Amount: \$569,469

Start Date: June 2019, Tentative End Date: May 2026

Investigator: **Shishir Chundawat** (Rutgers)

Agency: National Science Foundation (CBET - Cellular & Biochemical Engineering Program)

Role: PI

Project Title: *Computationally Designed Cellulases to Decrease the Cost of Biofuels*

Award Number: CSP-503631 Syn (DOE-JGI Community Science Program)

Grant Amount: Upto 500 kbp total DNA synthesis

Start Date: Sept 2018, Tentative End Date: Aug 2026

Investigators: **Shishir Chundawat** (Rutgers Lead PI), Jeffrey Linger (Note: Deanne Sammond was NREL PI before she left NREL in mid-2019)

Agency: Department of Energy (Joint Genome Institute Annual DNA Synthesis Award)

Role: PI

Expired Projects

Project Title: *Integrated conversion of cellulosic biomass and biomass-derived CO₂ to fuels and chemicals*

Award Number: S005193-USDA

Grant Amount: \$375,000 (not including \$160k total cost share contributions)

Start Date: Jan 2024, Tentative End Date: Sept 2025

Investigators: **Shishir Chundawat** (PI), Diane Hildebrandt (Co-PI), Karin Calvinho (Co-PI), Cornelius Masuku (Unfunded External Collaborator)

Agency: USDA (Northeast Sun Grant Program)

Role: PI

Project Title: *An Integrated Platform for Fully Automated, Continuous, & Real-Time Multi-Attributes Monitoring of Upstream Processes for Glycosylated Monoclonal Antibodies Production*

Award Number: PC5.2-112

Grant Amount: \$1,500,000 (including \$750k cost share for industry personnel time and donated equipment)

Start Date: Oct 2022, Tentative End Date: Mar 2025

Investigators: **Shishir Chundawat** (Lead PI), Oscar Potter (Co-I), Sean Gilliam (Co-I), Marianthi Ierapetritou (Co-I)

Agency: National Institute for Innovation in Manufacturing Biopharmaceuticals (NIIMBL)

Role: PI

Project Title: *Collaborative Research: Mechanism-guided enzyme engineering for fucosylated glycoconjugate synthesis*

Award Number: 1904862 (sub-award to Rutgers)

Grant Amount: \$182,223

Start Date: Aug 2022, Tentative End Date: Aug 2024

Investigators: **Shishir Chundawat** (Rutgers PI), Ronald Larson (University of Michigan PI)

Agency: National Science Foundation (CHE - Chemistry of Life Processes Program)

Role: Co-PI

Project Title: *Collaborative Research: Mechanism-guided enzyme engineering for fucosylated glycoconjugate synthesis*

Award Number: 1904890

Grant Amount: \$450,009

Start Date: Sept 2019, Tentative End Date: Aug 2024

Investigators: Heather Mayes (University of Michigan PI), **Shishir Chundawat** (Rutgers PI)

Agency: National Science Foundation (CHE - Chemistry of Life Processes Program)

Role: Co-PI

Project Title: *Understanding the structural features of reconstituted polysaccharide synthases*

Instrumentation: Cryo-EM Facility at EMSL

Award Number: 60012

Start Date: May 2021, Tentative End Date: Dec 2023

Investigator: **Shishir Chundawat** (Rutgers)

Agency: Pacific Northwest National Laboratory Cryo-EM/EMSL User Facility Access Grant

Role: PI

Project Title: *Understanding the role of inter-domain interactions of chimeric glycosyl hydrolases on the catalytic mechanism that enables transglycosylation versus hydrolysis reaction*

Instrumentation: Cryo-EM Facility at EMSL

Award Number: 60065

Start Date: May 2021, Tentative End Date: Dec 2023

Investigators: **Shishir Chundawat**, **Chandra Kanth Bandi** (Rutgers)

Agency: Pacific Northwest National Laboratory Cryo-EM/EMSL User Facility Access Grant

Role: PI

Project Title: *Research gift for Chundawat research program*

Award Number: -

Grant Amount: \$10,000 (Cash Gift) + \$36,455 (In-Kind Donation/Gift)

Start Date: Dec 2021, Tentative End Date: NA.

Investigator: **Shishir Chundawat** (Rutgers)

Agency: Croda Inc.

Role: PI

Project Title: *A Multiscale Approach to Characterizing Interfacial Carbohydrate-Active Enzymes*

Award Number: 1604421 (NSF-INTERN Supplement)

Grant Amount: \$54,816

Start Date: July 2021, Tentative End Date: Feb 2023

Investigator: **Shishir Chundawat** (Rutgers)

Agency: National Science Foundation (CBET – Catalysis Program)

Role: PI

Project Title: *A Multiscale Approach to Characterizing Interfacial Carbohydrate-Active Enzymes*

Award Number: 1604421 (NSF-INTERN Supplement)
Grant Amount: \$54,998
Start Date: July 2019, Tentative End Date: Feb 2023
Investigator: **Shishir Chundawat** (Rutgers)
Agency: National Science Foundation (CBET – Catalysis Program)
Role: PI

Project Title: *A Multiscale Approach to Characterizing Interfacial Carbohydrate-Active Enzymes*
Award Number: 1604421
Grant Amount: \$449,678
Start Date: Sept 2016, Tentative End Date: Feb 2023
Investigators: **Shishir Chundawat** (Rutgers Lead PI), Matthew Lang (Vanderbilt)
Agency: National Science Foundation (CBET – Catalysis Program)
Role: PI

Project Title: *In planta single-molecule imaging and holographic force spectroscopy to study real-time, multimodal turnover dynamics of polysaccharides and associated carbohydrate metabolites*
Award Number: DE-SC0019313
Grant Amount: \$1,500,000
Start Date: Sept 2018, Tentative End Date: Aug 2022
Investigators: Sanghyuk Lee (Rutgers Lead PI), **Shishir Chundawat** (Rutgers Lead Co-PI), Eric Lam, Matthew Lang (Vanderbilt), Wellington Muchero (ORNL), Sai Venkatesh Pingali (ORNL)
Agency: Department of Energy (Bioimaging Program in Biological & Environmental Research)
Role: Co-PI

Project Title: *Advanced continuous upstream manufacturing of biotherapeutics*
Award Number: 1R01FD006588
Grant Amount: \$1,800,000
Start Date: Sept 2018, Tentative End Date: Aug 2022
Investigators: Marianthi Ierapetritou (Original PI moved to U. Delaware) and **Shishir Chundawat** (Rutgers Lead PI), Rohit Ramachandran, Haoran Zhang, Georgios Tsilomelekis, Ravendra Singh, Doug Hausner
Agency: Food and Drug Administration
Role: Co-PI

Project Title: *Automated N-linked glycan analysis PAT to enable continuous biologics manufacturing*
Award Number: 4481
Grant Amount: \$63,439
Start Date: July 2020, Tentative End Date: Dec 2022.
Investigator: **Shishir Chundawat** (Rutgers)
Agency: Agilent Research Gift
Role: PI

Award Number: FP00010677 for Biacore 8K+ instrumentation

Start Date: Aug 2019, Tentative End Date: NA.

Investigators: Nilgun Tumer (PI), Study Team Member: **Shishir Chundawat** (along with several faculty)

Agency: National Institutes of Health (Shared Instrumentation Grant)

Role: Study Team Member

Project Title: *SusChEM-Designer Glycoligands for Enabling Targeted Multimodal Protein Bioseparations*

Award Number: 1704679

Grant Amount: \$300,000

Start Date: Sept 2017, Tentative End Date: Nov 2022

Investigator: **Shishir Chundawat** (Rutgers)

Agency: National Science Foundation (CBET - Separations Program)

Role: PI

Project Title: *In-vitro Multiscale Carbohydrate-Active Enzyme Binding Characterization using SANS*

Instrumentation: HFIR/BioSANS Small Angle Neutron Scattering at Oak Ridge National Laboratory

Award Number: 17752

Beam Time Date: 7-10 Aug 2017

Investigator: **Shishir Chundawat** (Rutgers)

Agency: Oak Ridge National Lab Neutron Sciences Division HFIR-BioSANS Instrumentation Grant

Role: PI

Project Title: *In-situ Time Resolved Ammonia-Salt Based Pretreatment of Cellulosic Biomass*

Instrumentation: HFIR/BioSANS Small Angle Neutron Scattering at Oak Ridge National Laboratory

Award Number: 17764

Beam Time Date: 7-10 Aug 2017

Investigator: **Shishir Chundawat** (Rutgers)

Agency: Oak Ridge National Lab Neutron Sciences Division HFIR-BioSANS Instrumentation Grant

Role: PI

Project Title: *Valorization of biopolymers to fuels, chemicals, & advanced materials*

Grant Amount: \$10,000

Start Date: May 2016, End Date: July 2017

Investigator: **Shishir Chundawat** (Rutgers)

Agency: Ralph E. Powe Junior Faculty Enhancement Award (ORAU)

Role: PI

Project Title: *Solving a Sticky Problem-Understanding Enzyme Binding to Lignocellulosic Biomass during Biofuel Production*

REU Supplement Subaward for Award Number: 1236120

Grant Amount: \$14,683

Start Date: Aug 2015, End Date: Dec 2016

Investigators: **Shishir Chundawat** (Rutgers PI), Timothy Whitehead (MSU PI)

Agency: National Science Foundation (CBET - Energy for Sustainability Program)

Role: Co-PI

Project Title: *Industrial Service Project with Aurobindo Pharma (NJ) on characterization of pharmaceutical formulations*

Grant Amount: \$1,975

Start Date: Dec 2015, End Date: Dec 2016

Investigator: **Shishir Chundawat** (Rutgers)

Agency: Aurobindo Pharma (NJ)

Role: PI

Project Title: *Solving a Sticky Problem: Understanding Enzyme Binding to Lignocellulosic Biomass during Biofuel Production*

Award Number: 1236120

Grant Amount: \$299,801

Start Date: Aug 2012, End Date: Dec 2016

Investigators: Timothy Whitehead (MSU), **Shishir Chundawat** (MSU and Rutgers)

Agency: National Science Foundation (CBET - Energy for Sustainability Program)

Role: Co-PI

Project Title: *Conversion of palm industry cellulosic waste to biofuels & biochemicals*

Award Number: MSU-DER #RC101539

Grant Amount: \$177,025

Start Date: 2012, Tentative End Date: 2014

Investigators: Venkatesh Balan, Bruce Dale, **Shishir Chundawat** (MSU)

Agency: Date Palm Research Center (King Faisal University)

Role: Co-PI

Internally Funded Research and User Facility Grants

Current Projects

Project Title: *High-throughput Protein/Therapeutics BioFoundry via Robotics and AI/ML/Data Science*

Start Date: Jan 2023, Tentative End Date: NA.

Investigators: Sagar Khare (Lead PI), **Shishir Chundawat**

Agency: Rutgers CCB

Role: Team Member/Collaborator

Project Title: *An AI-Enabled digital twin in bio-pharmaceutical manufacturing*

Grant Amount: \$25,000

Start Date: Feb 2024, Tentative End Date: Aug 2024.

Investigators: Rohit Ramachandran (PI), Weihong (Grace) Guo, Yuebin Guo, Adam Gormley, **Shishir Chundawat** (Rutgers)

Agency: Rutgers Chancellor Challenge-Cyberinfrastructure for AI in Science & Engineering (CASS)

Role: Team Member/Collaborator

Project Title: *Preserving and Nurturing Our Healthy Microbiome*
Start Date: Sept 2020, Tentative End Date: NA.
Investigators: Liping Zhao (Lead PI), **Shishir Chundawat** (along with 100+ Rutgers faculty)
Agency: Rutgers Big Idea Initiative
Role: Team Member/Collaborator

Expired Projects

Award Title: *A. Walter Tyson Assistant Professorship Endowment Award*
Grant Amount: \$12,500
Start Date: Jan 2022, End Date: June 2022.
Investigator: **Shishir Chundawat** (Rutgers)
Agency: Rutgers University School of Engineering (SOE) Dean's Office
Role: PI

Project Title: *Multimodal Single-molecule Microscopy (MoSoM) initiative to catalyze interdisciplinary life sciences research at Rutgers University*
Grant Amount: \$70,000
Start Date: Sept 2019, Tentative End Date: Aug 2022.
Investigators: Sang-Hyuk Lee, **Shishir Chundawat**
Agency: Rutgers Office of Research and Innovation Team Proposal Development Grant
Role: Co-PI

Project Title: *Responsive Bio-Inspired Materials for Sensing and Actuation*
Grant Amount: \$50,000
Start Date: Oct 2019, Tentative End Date: Sept 2020.
Investigators: Adam Gormley (PI), Vikas Nanda, Sagar Khare, Enver Izgu, **Shishir Chundawat**, Meenakshi Dutt, Benjamin Schuster (Rutgers)
Agency: Rutgers Initiative for Materials Research (iMR) Team Proposal Development Grant
Role: Team Member/Collaborator

Project Title: *Synthetic Cells and Cell Components*
Grant Amount: \$60,000
Start Date: Oct 2019, Tentative End Date: Sept 2020.
Investigators: Adam Gormley (PI), Vikas Nanda, Sagar Khare, Enver Izgu, **Shishir Chundawat**, Meenakshi Dutt, Benjamin Schuster (Rutgers)
Agency: Rutgers Office of Research and Innovation Team Proposal Development Grant
Role: Team Member/Collaborator

Project Title: *Enabling the creation of a sustainable and economically viable cellulosic biofuels industry in Brazil*
Grant Amount: \$8,000
Start Date: Feb 2016, End Date: Dec 2017.
Investigator: **Shishir Chundawat** (Rutgers)

Agency: Rutgers GAIA International Collaborative Research Grants for Tenure-Track Faculty
Role: PI

Teaching Experience

Doctoral Dissertation Primary Advisor at Rutgers

Current

1. **Ms. Samantha Shimabukuro** (2026-Current; Rutgers CBE Ph.D. Dissertation Title "*TBD*").
2. **Ms. Rebecca Garcia** (2025-Current; Rutgers CBE Ph.D. Dissertation Title "*TBD*").
3. **Mr. Austin Seamann** (2024-Current; Rutgers iQB Ph.D. Dissertation Title "*Harnessing Machine Learning for Protein Design: Advancing Sequence and Structure Design for Functional Applications*"). Co-advised with Sagar Khare (CCB/iQB).
4. **Mr. Ryan Murphy** (2024-Current; Rutgers CBE Ph.D. Dissertation Title "*TBD*").
5. **Ms. Srividya Vyjayanthi Tallavajhula** (2023-Current; Rutgers CCB Ph.D. Dissertation Title "*Development of high-throughput screening systems for carbohydrate active enzymes for fucosylated prebiotics synthesis*"). Co-advised with Sagar Khare (CCB/iQB).
6. **Mr. Antonio DeChellis** (2022-2026; Rutgers CBE Ph.D. Dissertation Title "*Charge Optimization of Hydrolase Enzymes and their Associated Binding Modules for Waste Polymer Degradation*").

Former

1. **Mr. Mohit Kumar** (2020-2025; Rutgers CBE Ph.D. Dissertation Title "[*Development of screening platforms for chemoenzymatic oligosaccharide synthesis*](#)").
2. **Mr. Dharanidaran Jayachandran** (2019-2023; Rutgers CBE Ph.D. Dissertation Title "[*A Multifaceted Bioengineering Toolbox to Enable In Vivo and In Vitro Characterization of Polysaccharide Synthases*](#)").
3. **Mr. Markus Hackl** (2018-2023; Rutgers CBE Ph.D. Dissertation Title "[*Development of a single-molecule force spectroscopy technique for the characterization of protein-carbohydrate dissociation forces*](#)").
4. **Mr. Aron Gyorgypal** (2019-2023; Rutgers CBE Ph.D. Dissertation Title "[*Enabling and investigating real-time monoclonal antibody N-linked glycosylation for upstream processing of biotherapeutics*](#)").
5. **Mr. Bhargava Nemmaru** (2017-2021; Rutgers CBE Ph.D. Dissertation Title "[*Cellulase Engineering to Alleviate Non-Productive Enzyme Binding to Pretreated Lignocellulosic Biomass*](#)").
6. **Mr. Chandra Kanth Bandi** (2016-2020; Rutgers CBE Ph.D. Dissertation Title "[*Novel Protein Engineering Approaches for Chemoenzymatic Synthesis of Glycans*](#)").

Master's Thesis Primary Advisor at Rutgers

Current

1. **Mr. Mitun Aghipothu Umashankar** (2025-Present; Rutgers CBE MS Thesis Title "*TBD*")

Former

1. **Mr. Antonio De Chellis** (2020-2022; Rutgers CBE MS Thesis Title "[*Supercharging of a family-5 glycoside hydrolase Cel5A from Thermobifida fusca to improve cellulose hydrolysis*](#)")

2. **Mr. Antash Chaturvedi** (2019-2022; Rutgers CBE MS Thesis Title "[*Investigation of process conditions and chemical modulators on CHO-cell based trastuzumab production*](#)")
3. **Mr. Zachary Power** (2019-2021; Rutgers CBE MS Thesis Title "[*Covalent linkage of cello-oligosaccharides to borosilicate glass for single molecule binding measurements using acoustic force spectroscopy*](#)")
4. **Ms. Madhura Kasture** (2017-2020; Rutgers CBE MS Thesis Title "[*Impact of carbohydrate binding modules on transglycosylation efficiency in the glycoside hydrolase family 5 proteins*](#)")
5. **Mr. Antonio Goncalves** (2017-2019; Rutgers CBE MS Thesis Title "[*Chemoenzymatic synthesis of multimodal glycoligands with bio-orthogonal aldehyde based functional moiety*](#)")
6. **Ms. Ayushi Agrawal** (2016-2019; Rutgers CBE MS Thesis Title "[*Development of a click chemistry based screening methodology for the directed evolution of glycosynthases*](#)")
7. **Ms. Namratha Subhash** (2016-2018; Rutgers CBE MS Thesis Title "[*Selective growth of targeted bacteria using enzymatically synthesized oligosaccharides*](#)")
8. **Mr. Akash Dagia** (2016-2018; Rutgers CBE MS Thesis Title "[*Impact of a conserved tyrosine residue on binding of family 1 carbohydrate binding modules to cellulose allomorphs*](#)")
9. **Ms. Yuxin Liu** (2015-2017; Rutgers CBE MS Thesis Title "[*Characterization of Thermobifida fusca cellulase activity on unnatural cellulose allomorphs*](#)")
10. **Ms. Vibha Narayanan** (2015-2017; Rutgers CBE MS Thesis Title "[*Binding interactions of Family 1 CBMs with cellulose allomorphs*](#)")

Master's Thesis Committee Member at Rutgers

Former

1. **Mr. Pankaj Jadhav** (Oct 2023; Rutgers CBE MS Thesis and Defense Committee Member "[*Biomanufacturing of encapsulated mesenchymal stem cells in a vertical wheel bioreactor*](#)"; MS Thesis Director - Dr. Biju Parekkadan)
2. **Mr. Aparajith Bhaskar** (Oct 2018; Rutgers CBE MS Thesis and Defense Committee Member "[*Implementation of an Advanced Control Strategy into a Continuous Direct Compaction Pharmaceutical Tablet Manufacturing Process*](#)"; MS Thesis Director - Dr. Ravendra Singh)
3. **Mr. Praneeth Annam** (Jan 2018; Rutgers CBE MS Thesis and Defense Committee Member "[*Life Cycle Assessment for Bio-Based Production of p-Xylene*](#)"; MS Thesis Director - Dr. Marianthi Ierapetritou)
4. **Mr. Geetartha Uppaladadi** (May 2016; Rutgers CBE MS Thesis and Defense Committee Member "[*Mesoscale modeling of biomimetic macromolecular aggregates*](#)"; MS Thesis Director - Dr. Meenakshi Dutt)

Postdoctoral Trainees at Rutgers

Current

1. **Dr. Aditya Narvekar** (March 2023-Current; Full-time Postdoctoral Research Associate at Rutgers University). Project title: *Real-time analytics for glycosylated biologics manufacturing*.

Former

1. **Dr. Chandra Kanth Bandi** (May 2021-Jan 2023; Full-time Postdoctoral Research Associate at Rutgers University). Project title: *CAZyme engineering for human milk oligosaccharides synthesis*.

2. **Dr. Viki Chopda** (May 2019-Dec 2020; Full-time Postdoctoral Research Associate at Rutgers University). Project title: *Continuous biomanufacturing of glycosylated monoclonal antibodies*.
3. **Dr. Mohammad Irfan** (Feb 2019-Nov 2020; Full-time Postdoctoral Research Associate at Rutgers University). Project title: *Heterologous expression of polysaccharide synthases in plant protoplasts*.
4. **Dr. Chao Zhao** (Aug 2018-July 2019; Visiting Postdoctoral Scholar from China; Currently a faculty member at Zhejiang A&F University).
5. **Dr. Shyamal Roy** (Jul 2016-June 2017; Visiting Postdoctoral Scholar from India; Currently a faculty member at Jadavpur University)
6. **Dr. Yongling Qin** (Jan 2016-Dec 2016; Visiting Postdoctoral Scholar from China; Currently a faculty member at Hechi University)

Students Supervised for Independent Studies at Rutgers

Graduate Students

1. **Mr. Rahul Thakur** (Aug 2024-Jan 2026; Rutgers CBE MS Independent Study Project Title "*Glycobiologics production via continuous microperfusion with integrated glycosylation monitoring using N-GLYcanalyzer PAT*")
2. **Mr. Gautam Reddy Tupalle** (Sept 2025-Apr 2026; Rutgers CBE MS Independent Study Project Title "*Conversion of dairy industry waste sugars into Human Milk Oligosaccharides*")
3. **Ms. Nurulhuda Binte Suaini** (Sept 2024-Sept 2025; Research Intern from University of Surrey (UK); Project Title "*PAT for Advanced GlycoBiologics Manufacturing*")
4. **Mr. Rishabh Shah** (Aug 2021-Dec 2022; Rutgers CBE MS Independent Study Project Title "*Glycoproteins production and characterization to enable biologics manufacturing*")
5. **Mr. Robert Kearney Jr** (Dec 2021-June 2022; Rutgers BME ME Independent Study Project Title "*Expression, Reconstitution, and Characterization of Hyaluronan Synthase Enzymes*")
6. **Ms. Erin K. Wilson** (Summer 2021-Dec 2021; Rutgers BME ME Independent Study Project Title "*Recent Advances in Preparation Techniques and Analytical Methods to Support Rapid N-Glycan Analysis of Monoclonal Antibodies (mAbs)*")
7. **Ms. Margaux Thiry** (2019 Summer; University of Poitiers-France Visiting Biology MS Student (Biochemistry Molecular and Cell Biology and Genetics Program), Summer Intern at Rutgers). Internship Project/Report Titled "*Operon Optimization for Bacterial Protein Expression*" was submitted to University of Poitiers for credits.
8. **Ms. Navya Sahithi** (2016-2019; Rutgers CBE MS). Trained in lab and conducted research on NSF Sponsored Project (Award #1604421) to express supercharged CAZymes and characterize their binding affinity to cellulose/lignin. Submitted a final independent study report to CBE titled "*Characterizing binding and activity of supercharged CAZymes to lignocellulosic biomass*" in Summer 2019.
9. **Mr. Bolun Tan** (2016-2018; Rutgers CBE MS). Trained in lab and conducted research on Aspen Plus modeling project. Submitted a final independent study report to CBE titled "*A solid liquid equilibrium based on eNRTL(electrolyte NRTL) model and evaluation of thermodynamic representation of NH₄SCN/NH₃/Ethanol system*".
10. **Ms. Shraddha Gupta** (2016-2018; Rutgers CBE MS). Trained in lab and conducted research on NSF Sponsored Project (Award #1704679) to express proteins and characterize their binding affinity.

Submitted a final independent study report to CBE on glycoligand-beads based protein bioseparations.

11. **Mr. Shashwat Gupta** (2015-2016, Rutgers CBE MS); Trained in lab and conducted research on ORAU Sponsored Project to characterize ammonia pretreated biomass and its enzymatic digestibility.
12. **Mr. Satvik Sharma** (2015-2016; Rutgers CBE MS); Trained in lab and conducted research on NASA Sponsored Project (New Jersey Space Grant Consortium Graduate Student Fellowship) to test activity of CAZymes under various mixing regimes.
13. **Mr. Soumya Asthana** (2015-2016; Rutgers CBE MS); Trained in lab and conducted research on NSF Sponsored Project (Award #1236120) to express supercharged CAZymes and characterize their binding affinity to lignin.

Undergraduate Students

1. **Ms. Maya Thakrar** (2025-Present; Rutgers CBE BS; Aresty Summer Science); Trained in lab and conducted research protein expression and characterization for seaweed degradation and presented research at 2026-2027 Aresty Research Symposium.
2. **Mr. Jayveer Rathod** (2026-Present; Rutgers CBE BS); Training in lab and conducting research on protein expression and characterization for seaweed degradation.
3. **Mr. Brian Ho** (2026-Present; Rutgers CBE BS); Training in lab and conducting research on protein expression and characterization for seaweed degradation.
4. **Ms. Sarah Coven** (2026-Present; Rutgers CBE BS); Training in lab and conducting research on protein expression and characterization for seaweed degradation.
5. **Ms. Raadha Garg** (2025-Present; Rutgers CBE BS; Rutgers Douglass WiSE SUPER Program); Training in lab and conducting research on protein expression and characterization for seaweed degradation. Presented research work at SUPER summer program studying effect of CAZyme engineering for seaweeds degradation.
6. **Ms. Merlyn Roy** (2025-Present; Rutgers CBE BS; Rutgers Douglass WiSE SUPER Program); Training in lab and conducting research on protein expression and characterization for single-molecule protein-glycan interactions. Presented research work at SUPER summer program studying effect of CAZyme engineering for single-molecule protein-glycan interactions.
7. **Ms. Sofia Colella** (2025-Present; Rutgers CBE BS; Rutgers Douglass WiSE SUPER Program); Training in lab and conducting research on protein expression and characterization for single-molecule protein-glycan interactions. Presented research work at SUPER summer program studying effect of CAZyme engineering for single-molecule protein-glycan interactions.
8. **Ms. Shreyoshi Ghosh** (2025-Present; Rutgers CBE BS); Training in lab and conducting research on protein expression and characterization for single-molecule protein-glycan interactions.
9. **Ms. Lara Erdogan** (2025-Present; Rutgers CBE BS; Federal Workstudy Research Assistant); Training in lab and conducting research on protein expression and characterization for seaweed degradation.
10. **Ms. Yaro Varnichenko** (2026-Present; Rutgers CBE & Biological Sciences BS); Training in lab and conducting research on protein expression and characterization for HMOs synthesis.
11. **Mr. Judah Anyobodeh** (Spring 2026-Present; Rutgers Biological Sciences BS); Training in lab and conducting research on protein expression and characterization. Submitted CBN 119:307-308

independent study report "*Using Whey as an Alternative Feedstock for Microbial Production of HMOs*" for research credits in Spring 2026.

12. **Mr. Giovanni Pescatore** (Spring 2026; Rutgers CBE BS); Training in lab and conducting research on continuous glycosylated antibody production and characterization.
13. **Mr. Malcolm Harrison** (2025-2026; Rutgers CBE BS); Training in lab and conducting research on continuous glycosylated antibody production and characterization.
14. **Ms. Padmadisha Khot** (2025-Present; Rutgers CBE BS); Training in lab and conducting research on glycosylated antibody production and characterization. Submitted CBE 14:155:391-392 independent study report "*Advanced manufacturing of glycosylated monoclonal antibodies*" for research credits in Spring 2026.
15. **Ms. Elayna Salle** (2025-Present; Rutgers CBE BS); Training in lab and conducting research on protein expression and characterization for HMO synthesis. Submitted CBE 14:155:391-392 independent study report "*Engineering E. coli for the Production of 2'-Fucosyllactose from Recycled Whey Lactose*" for research credits in Fall 2025 and Spring 2026.
16. **Ms. Ashwika Chandu** (2025-Present; Rutgers Public Health BS); Training in lab and conducting research on protein expression and characterization for HMOs synthesis from seaweed wastes.
17. **Ms. Pauline Semira** (2025-Present; Rutgers CBE BS); Training in lab and conducting research on glycosylated antibody production and characterization. Submitted CBE 14:155:391-392 independent study report "*Extraction and Characterization Of Exosomes Isolated From Chinese Hamster Ovary Cells*" for research credits in Fall 2025 and Spring 2026.
18. **Ms. Srivarshini Ganapathyappan Muthusamy** (2025-Present; Rutgers BME BS); Training in lab and conducting research on protein expression and characterization. Submitted BME 14:125:291-292 independent study report "*Glycoengineering of dairy waste sugars into Human Milk Oligosaccharides*" for research credits in Spring 2025.
19. **Ms. Akshanth Padmanabhan** (2025-Present; Rutgers CBE BS); Training in lab and conducting research on glycosylated antibody production and characterization. Submitted CBE 14:155:291-292 independent study report "*Advanced manufacturing of glycobiologics*" for research credits in Fall 2025.
20. **Mr. Sumay Trivedi** (2024-2026; Rutgers CBE BS); Training in lab and conducting research on enzyme engineering and characterization for cellulose degradation.
21. **Ms. Kendra Marie Buffaloe** (2024-2025; Rutgers BME BS); Training in lab and conducting research on NIIMBL Sponsored project focused on mammalian cell culture and glycobiologics characterization analytics.
22. **Ms. Sophia Fernandez** (2024-2025; Rutgers CBE BS; Rutgers Douglass WiSE SUPER Program); Training in lab and conducting research on protein expression and characterization for NSF HMO project. Presented research work at SUPER summer program studying effect of CAZyme engineering to produce HMO synthesizing glycosynthases.
23. **Ms. Arushi Biswas** (2024-2025; Rutgers CBE BS; Rutgers Douglass WiSE SUPER Program); Training in lab and conducting research on protein expression and characterization for NSF HMO project. Presented research work at SUPER summer program studying effect of CAZyme engineering to produce HMO synthesizing glycosynthases.
24. **Ms. Mahati Salapu** (2024-2025; Rutgers CBE BS; Rutgers Douglass WiSE SUPER Program); Training in lab and conducting research on protein expression and characterization for NSF CAREER project.

Presented research work at SUPER summer program studying effect of protein supercharging on binding to cellulose using acoustic force spectroscopy.

25. **Ms. Shreya Dhillon** (2024-2025; Rutgers CBE BS); Training in lab and conducting research on protein expression and characterization for NSF CAREER project.
26. **Ms. Iris Brody** (2024-2025; Rutgers CBE BS; Rutgers Douglass WiSE SUPER Program); Training in lab and conducting research on NIIMBL Sponsored project focused on mammalian cell culture and glycobionics characterization analytics. Presented research work at SUPER summer program studying effect of mannose addition on CHO cell glycosylation pathways.
27. **Ms. Nabanita Paul** (2024 Summer; Indian Institute of Technology UG Student (Biosciences and Bioengineering Program), Summer Intern at Rutgers). Internship Projects/Reports Titled "*Development of High-throughput Screening Systems for Glycosynthase enzymes using a Dual Plasmid Strategy*" was submitted to Khorana Scholar Program.
28. **Ms. Samantha Shimabukuro** (2023-2025; Rutgers CBE BS; Slade Program); Trained in lab and conducted research on to characterize CAZymes involved in glycans/plastics degradation.
29. **Mr. Samir Verma** (2023-2024; Rutgers CBE BS; Research Assistant); Training in lab and conducting research on NIIMBL Sponsored project focused on mammalian cell culture and glycobionics characterization analytics.
30. **Ms. Parbeen Kaur Swaich** (2023-2024; Rutgers BME BS; Federal Workstudy Research Assistant); Training in lab and conducting research on NIIMBL Sponsored project focused on mammalian cell culture and glycobionics characterization analytics.
31. **Mr. Vincent Gambino** (2023-2024; Rutgers CBE BS; CBE Research Assistant); Trained in lab and conducted research to express, purify and characterize carbohydrate-active enzymes. Also, as part of CBE independent study, developed a granular solids mixing activity using a vibrating fluidizer for outreach events.
32. **Ms. Nakeitha Brown** (2023-2024; Post-Baccalaureate Research Experience for LSAMP Students (PRELS), Rutgers-Newark). PRELS scholar is working on project focused on "*Advanced Analytical Analysis of mAb N-Glycans*" and is supported for a year-long fellowship by the GS-LAMP Program.
33. **Mr. Gokulnath Ganesan** (2023 Summer; Indian Institute of Technology UG Student (Chemical Engineering Program), Summer Intern at Rutgers). Internship Projects/Reports Titled "*Rupture Force Determination of Single Point Mutant Carbohydrate-Binding Modules through the Lens of Acoustic Force Spectroscopy*" and "*Standardizing Automated Analysis of mAb-bound N-Glycans using Sequential Injection Analysis*" was submitted to Khorana Scholar Program.
34. **Mr. Tong Zhong** (Fall 2022/Spring 2023; Rutgers Biochemistry BS); Trained in lab and conducted independent study research work as part of Rutgers SEBS Research Problems in Biochemistry Course (11:115:493/494). Submitted report titled "*In vivo Biotinylation of Carbohydrate-Binding Proteins*" to Rutgers SEBS Programs for Credit in Fall 2022/Spring 2023.
35. **Mr. Yossef Fakry** (Fall 2022; Rutgers Microbiology BS); Trained in lab and conducted independent study research work as part of Rutgers SEBS Research in Microbiology Course (11:680:497/498). Submitted report titled "*Enzyme engineering for fucosylated human milk oligosaccharides biosynthesis*" to Rutgers SEBS Programs for Credit in Fall 2022.
36. **Ms. Brittany Burke** (Fall 2021; Rutgers SEBS BS); Trained in lab and conducted independent study research work as part of School of Environmental & Biological Sciences General (four-year) Honors Program Tutorial III Program. Submitted report on "*Enzyme engineering for fucosylated human milk oligosaccharides biosynthesis*" to Rutgers SEBS Honors Programs for Credit in Fall 2021.

37. **Mr. Edward Contrada** (2019-2021; Rutgers BME BS; Aresty Research Assistant); Trained in lab and conducted research on Rutgers Sponsored project titled '*Developing an Acoustic Force Spectroscopy Technique to Measure Carbohydrate Binding Module Interaction with Cellulose*'. Research presented at 2020 Aresty Research Symposium. Submitted report on "*Measuring DNA tether binding efficiency using Pico-Green dye*" to Rutgers CBE Programs for Credit in Fall 2020-Spring 2021.
38. **Mr. Khovesh Ramdin** (2021-2022; Rutgers Physics BS); Training in lab and conducting research on NSF Sponsored Project (Award #1846797) to develop MATLAB code for single-molecule force spectroscopy experiments data analysis and simulation of Tethered Particle Motion for educational purposes. Research to be published in peer-reviewed journal.
39. **Mr. Srivatsan Shankar** (2020-2021; Rutgers CBE BS; Aresty Research Assistant); Trained in lab and conducted research on Rutgers Sponsored project titled '*Studying the effect of CBM supercharging on endo-exo enzyme synergism on lignocellulosic biomass*'. Research presented at 2021 Aresty Research Symposium. Submitted report to Rutgers CBE Program for Credit in Spring 2021.
40. **Mr. Kishan Mehta** (2020-2021; Rutgers CBN BS; Aresty Undergraduate Research Fellow); Received Aresty Undergraduate Research Award to conduct research on DOE Sponsored Project (Award No. DE-SC0019313) to engineer, express and purify plant membrane proteins for imaging in vitro.
41. **Mr. Shamanth Manjunatharao** (2020-2021; Rutgers Chem BS; Aresty Undergraduate Research Fellow); Received Aresty Undergraduate Research Award to conduct research on DOE Sponsored Project (Award No. DE-SC0019313) to engineer, express and purify plant membrane proteins for imaging in vitro.
42. **Mr. Tan Ngo** (2020-2021; Rutgers CE BS; Aresty Research Assistant); Training in lab and conducting research on NSF Sponsored Project (Award #1846797) to develop LabView code for single-molecule force spectroscopy experiments data analysis. Research titled '*Dynamic simulation of DNA-tethered particle motion*' to be presented in Aresty 2021 Symposium.
43. **Mr. Kevin Zheng** (2020-2021; Rutgers CBE BS; Aresty Research Assistant); Training in lab and conducting research on NSF Sponsored project (Award No. 1904890) titled '*Enzyme engineering for synthesis of prebiotic human milk oligosaccharides*'. Research to be presented at 2021 Aresty Research Symposium.
44. **Mr. Igor Guranovic** (2020-2023; Rutgers BME BS; Aresty Research Assistant); Training in lab and conducting research on NSF Sponsored project (Award No. 1904890) titled '*Enzyme engineering for synthesis of prebiotic human milk oligosaccharides*'. Research to be presented at 2021 Aresty Research Symposium.
45. **Mr. Vrushabh Khot** (2020-2021; Rutgers CBE BS; Aresty Research Assistant); Training in lab and conducting research on FDA Sponsored project (Award No. 5R01FD006588) titled '*Glycosylated monoclonal antibody production and mAbs characterization for enabling Quality-by-Control biomanufacturing*'. Research to be presented at 2021 Aresty Research Symposium.
46. **Mr. Rishabh Shah** (2020-2023; Rutgers CBE BS; Aresty Research Assistant); Training in lab and conducting research on FDA Sponsored project (Award No. 5R01FD006588) titled '*Glycosylated monoclonal antibody production and mAbs characterization for enabling Quality-by-Control biomanufacturing*'. Research to be presented at 2021 Aresty Research Symposium.
47. **Ms. Vrinda Jain** (2020-2021; Rutgers BME BS; Aresty Research Assistant); Training in lab and conducting research on NSF Sponsored project (Award No. 1904890) titled '*Enzyme engineering for*

synthesis of prebiotic human milk oligosaccharides'. Research to be presented at 2021 Aresty Research Symposium.

48. **Mr. Darin Mak** (2020-2021; Rutgers BME BS; Aresty Research Assistant); Training in lab and conducting research on NSF Sponsored project (Award No. 1904890) titled '*Enzyme engineering for synthesis of prebiotic human milk oligosaccharides*'. Research to be presented at 2021 Aresty Research Symposium.
49. **Ms. Shoili Banerjee** (2020-2023; Rutgers CBE BS; Rutgers Energy Institute or REI Undergraduate Fellow); Trained in lab and conducted research on REI and DOE Sponsored Project (Award #DE-SC0019313) to express, purify, and characterize mutant CAZymes. Student submitted online poster in spring 2020 titled 'Expressing Cesa8 and Cesa5 in *P. pastoris* for in vitro cellulose analysis' for DRC course on Intro to Scientific Research. Final student report on "*Expression, purification and characterization of a plant origin cellulose synthase enzyme*" to be presented and submitted to REI in coming 2020-2021 academic year.
50. **Ms. Vaneeza Abbas** (2019-2020; Rutgers CBE BS; CBE Research Assistant); Trained in lab and conducted research on development of reproducible surface modification procedures for the attachment of glycans and proteins to study protein-glycan interactions at the single molecule level.
51. **Mr. Atharv Kulkarni** (2019-2020; Rutgers BME BS; Aresty Research Assistant); Developed LabView code for single-molecule force spectroscopy experiments data analysis relevant to NSF Sponsored Project. Research presented at 2020 Aresty Research Symposium.
52. **Ms. Wen-Chen Chen** (2019-2021; Rutgers CBE BS; CBE Research Assistant); Trained in lab and conducted research on NSF Sponsored Project (Award #1704679) to express, purify and characterize engineered proteins and synthesized glycans. Submitted CBE independent study report for research credits in Fall 2019/Spring 2020.
53. **Ms. Jenna Douglas** (2019-2023; Rutgers CBE BS; Aresty and CBE Research Assistant); Trained in lab and conducted research on NSF Sponsored Project (Award #1604421) to express, purify and characterize processive cellulase/CBMs.
54. **Mr. Naven Sekhon** (2019-2020; Rutgers CBE BS; Aresty and CBE Research Assistant); Trained in lab and conducted research on NSF Sponsored Project (Award #1604421) to express, purify and characterize processive cellulase/CBMs.
55. **Mr. Ahmed Abdelhamid** (2019-2020; Rutgers BME BS; Aresty Undergraduate Research Fellow); Received Aresty Undergraduate Research Award to conduct research on DOE Sponsored Project (Award No. DE-SC0019313) to engineer, express and purify plant membrane proteins for imaging in vitro.
56. **Mr. Srivatsan Shankar** (2019-2021; Rutgers CBE BS; Aresty and CBE Research Assistant); Trained in lab and conducted research on NSF Sponsored Project (Award #1604421) to express, purify and characterize processive cellulase/CBMs.
57. **Ms. Deepika Saravana** (2019-2020; Rutgers CBE BS; Aresty Research Assistant); Trained in lab and conducted research on NSF Sponsored Project (Award #1704679) to optimize column chromatography technique for preparative sugar separations and characterize enzymatic reaction for glycan synthesis. Will be presenting a poster at Aresty Symposium 2020.
58. **Ms. Khushbu Patel** (2019-2020; Rutgers CBE BS; Aresty Research Assistant); Trained in lab and conducted research on NSF Sponsored Project (Award #1704679) to express, purify and

characterize engineered proteins and synthesized glycans. Will be presenting a poster at Aresty Symposium 2020.

59. **Mr. Jorge Gustavo Tapia** (2019-2020; Rutgers CBE BS; Aresty Research Assistant); Trained in lab and conducted research on NSF Sponsored Project (Award #1704679) to express, purify and characterize supercharged green fluorescent proteins (GFP) to test their binding characteristics to sugar ligands. Will be presenting a poster at Aresty Symposium 2020.
60. **Mr. Akshay Thakur** (2019-2020; Rutgers BT BS; Aresty Undergraduate Research Fellow); Received Aresty Undergraduate Research Award to conduct research on DOE Sponsored Project (Award No. DE-SC0019313) to engineer, express and purify plant membrane proteins for imaging in vitro.
61. **Mr. Ari Alter** (2019-2020; Rutgers CBE BS; Research Assistant); Trained in lab and conducting research on FDA Sponsored project (Award No. 5R01FD006588) titled '*Glycosylated monoclonal antibody production and mAbs characterization for enabling Quality-by-Control biomanufacturing*' developing models for Raman PAT data chemometrics.
62. **Ms. Rana Said** (2019-2021; Rutgers Pharmacy BPharmD; Research Assistant); Trained in lab and conducting research on FDA Sponsored project (Award No. 5R01FD006588) titled '*Glycosylated monoclonal antibody production and mAbs characterization for enabling Quality-by-Control biomanufacturing*' on PNGase enzyme heterologous expression.
63. **Ms. Sarah Hussain** (2019-2020; Rutgers CBE BS; CBE Research Assistant); Trained in lab and conducted research on NSF Sponsored Project (Award #1604421) to express, purify and characterize engineered proteins. Submitted CBE independent study report "*Supercharging of Cel6B, Cel5A, GFP, and CelE*" for research credits in Fall 2019/Spring 2020.
64. **Mr. Jonathan Ash** (Summer/Fall 2019; Rutgers Biotechnology (Bioinformatics) BS); Trained in lab and conducted independent study research work as part of School of Environmental & Biological Sciences General (four-year) Honors Program Tutorial III Program. Submitted report titled "*Acoustic force spectroscopy (AFS) Lab Research Report*" to Rutgers SEBS Honors Programs for Credit in Fall 2019. Submitted report titled '*In-silico Docking and Rupture Force Determination of Glycan Binding Protein Complexes*' to Rutgers SEBS George H. Cook Scholars Program for Credit in Fall 2020/Spring 2021.
65. **Ms. Lisa Poponne** (2018-2019; Rutgers CBE BS; Aresty and CBE Research Assistant); Trained in lab and conducted research on NSF Sponsored Project (Award #1604421) to express, purify and characterize processive cellulase/CBMs. Submitted CBE independent study report "*Production and Characterization of CBM Family 1 Aromatic Mutants for Single Molecule activity*".
66. **Mr. Mohamed Khan** (Spring 2019; Rutgers Biochemistry and Microbiology BS); Trained in lab and conducted independent study research work as part of Research in Biochemistry 11:115:494 Course. Submitted report titled "*Impact of planar aromatic residues of CBM on endo-cellulase activity*" to Rutgers Department of Biochemistry and Microbiology for Credit.
67. **Ms. Orchid Poponne** (2018-2019; Rutgers CBE BS; Aresty and CBE Research Assistant); Trained in lab and conducted research on NSF Sponsored Project (Award #1604421) to express, purify and characterize processive cellulase/CBMs. Submitted CBE independent study report "*The effect of aromatic residues on the activity of the Cel6B Catalytic Domain*".
68. **Ms. Ruchi Pandya** (2018-2019; Rutgers Biotechnology BS); Trained in lab and conducted independent study research work as part of School of Environmental & Biological Sciences Program. Submitted report titled "*Studying the effect of CBM supercharging on binding to cellulose*".

allomorphs" to Rutgers SEBS Research in Biotechnology Program in Fall 2018/Spring 2019 for Credit.

69. **Ms. Siqi Ma** (2018-2019; Rutgers Molecular Biology & Biochemistry BS; Aresty Summer Science); Trained in lab and conducted research on Rutgers Sponsored Project titled '*Developing an Acoustic Force Spectroscopy Technique to Measure Carbohydrate Binding Module Interaction with Cellulose*' and research presented at 2018 Aresty Research Symposium.
70. **Mr. Mohamed Khan** (Fall 2018; Rutgers Biochemistry and Microbiology BS); Trained in lab and conducted independent study research work as part of Research in Biochemistry 11:115:493 Course. Submitted report titled "*Impact of planar aromatic residues of CBM on exo-cellulase activity*" to Rutgers Department of Biochemistry and Microbiology for Credit.
71. **Ms. Jia-Mei Hong** (2018-2019; Rutgers CBE BS; Aresty Research Assistant); Trained in lab and developed cellulosic bioenergy outreach protocol as part of NSF Sponsored Project (Award #1604421) and assisted Dr. Chundawat in a summer 2018 high-school students outreach event sponsored by the Rutgers SOE.
72. **Mr. Allan Wang** (2018-2020; Rutgers CBE BS; Aresty Summer Science and Rutgers Energy Institute or REI Undergraduate Fellow); Trained in lab and conducted research on REI and NSF Sponsored Project (Award #1604421) to express, purify, and characterize mutant CAZymes. Final student report on "*Studying the effect of CBM supercharging on exo-cellulase activity: Optimization of supercharged mutant expression and purification*" presented and submitted to REI.
73. **Mr. Varun Raghuraman** (2017-2020; Rutgers CS BS; Aresty Summer Science & Undergraduate Research Fellow); Trained in lab and conducted research on NSF Sponsored Project (Award #1604421) to express, purify, and characterize mutant CAZymes and CBMs in 2017-2018. Received Aresty Undergraduate Research Award in 2019 to develop LabView code for single-molecule force spectroscopy experiments data analysis relevant to NSF Sponsored Project (Award #1846797).
74. **Mr. Benjamin Esposito** (2017-2018; Rutgers CBE BS; Aresty Research Assistant); Trained in lab and conducted research on cellulosic biomass composition analysis and enzymatic hydrolysis
75. **Mr. Patrick Doran** (2016-2018; Rutgers BME BS; Aresty Research Assistant); Trained in lab and conducted research on NSF Sponsored Project (Award #1604421) to model protein structure using Rosetta/FoldIt software package
76. **Ms. Marissa Berger** (2015-2018; Rutgers BME BS/MS; Aresty Researcher/Aresty Summer Science, NSF REU Scholar); Trained in lab and conducted research on NSF REU Sponsored Project (Award #1236120) to express, purify, and characterize CAZymes binding/activity
77. **Mr. Neelan Sivaneri** (2017-2020; Rutgers CBE BS; Aresty and CBE Research Assistant); Trained in lab and conducted research on NSF Sponsored Projects (Award #1604421 and #1704679) to express, purify and characterize CAZymes. Submitted CBE independent study report in Fall 2019.
78. **Mr. Zheyu Zhang** (2016-2017; Rutgers CBE BS; Aresty Research Assistant); Trained in lab and conducted research on characterizing protein adsorption to oxidized polysaccharide surfaces
79. **Ms. Elizabeth McGinley** (2016-2017; Rutgers CBE BS; Aresty Research Assistant & Dr. Alkis Constantinides Endowed Scholar); Trained in lab and conducted research on NSF Sponsored Project (Award #1236120) to express and purify CAZymes
80. **Ms. Cindy Farino** (2015-2017; Rutgers BME BS; Aresty Research Assistant & NSF REU Scholar); Trained in lab and conducted research on NSF REU Sponsored Project (Award #1236120) to express, purify, and characterize CAZymes binding to cellulose allomorphs

81. **Ms. Jihyun Park** (2015-2017; Rutgers CBE BS; Aresty Research Assistant & Dr. Alkis Constantinides Endowed Scholar); Trained in lab and conducted research on NSF Sponsored Project (Award #1236120) to express and purify CAZymes
82. **Ms. Alina Thokkadam** (2016-2019; Rutgers CBE BS; Aresty Summer Science); Trained in lab and conducted research on NSF Sponsored Project (Award #1704679) to characterize CAZymes involved in glycans synthesis
83. **Ms. Samantha Cobos** (Summer 2016; NSF REU Visiting Undergraduate Student from Pace University); Trained in lab and conducted research on NSF-REU Sponsored Project to characterize protein adsorption to oxidized polysaccharide surfaces
84. **Ms. Jeong Eunsu** (2015-2016; Rutgers CBE BS/MS); Conducted research on using Aspen Plus process simulation software to model cellulosic ethanol biorefinery using NREL base model as group project following completion of 155:324 design course
85. **Mr. Ashav Desai** (2015-2016; Rutgers CBE BS); Conducted research on using Aspen Plus process simulation software to model cellulosic ethanol biorefinery using NREL base model as group project following completion of 155:324 design course
86. **Mr. Alexander Pelham-Webb** (2015-2016; Rutgers CBE BS); Conducted research on using Aspen Plus process simulation software to model cellulosic ethanol biorefinery using NREL base model as group project following completion of 155:324 design course
87. **Ms. Sanchari Ghosh** (2015-2016; Rutgers CBE BS); Conducted research on using Aspen Plus process simulation software to model cellulosic ethanol biorefinery using NREL base model as group project following completion of 155:324 design course
88. **Mr. Raymond Bertram** (2015-2016; Rutgers CBE BS); Conducted research on using Aspen Plus process simulation software to model cellulosic ethanol biorefinery using NREL base model as group project following completion of 155:324 design course
89. **Mr. Jonathan Gerszberg** (2015-2016; Rutgers CBE BS); Developed basic hidden Markov type models for CAZymes
90. **Mr. Yianni Antonatos** (2015-2016; Rutgers CBE BS); Trained in lab on basics of microbiology, molecular biology, and enzymology
91. **Mr. Dylan McHugh** (2015-2016; Rutgers CBE BS); Trained in lab and conducted research on kinetic modeling of data generated via confocal fluorescence microscopy imaging of CAZyme desorption from cellulose microfibril surfaces
92. **Mr. Apurva Srivastav** (2015-2016; Rutgers CBE BS; Aresty Research Assistant); Trained in lab and conducted research on using Aspen Plus process simulation software to model cellulosic biomass ammonia pretreatment
93. **Ms. Maytal Merhav** (2015-2016; Rutgers CBE BS; Aresty Research Assistant); Trained in lab and conducted research on NSF Sponsored Project (Award #1236120) to express supercharged CAZymes and characterize their binding affinity to lignin
94. **Ms. Danielle Mainardi** (2015-2016; Rutgers CBE BS; Aresty Research Assistant); Trained in lab and conducted research on NSF Sponsored Project (Award #1236120) to express supercharged CAZymes and characterize their binding affinity to lignin
95. **Ms. Natália Dias** (Summer 2016; Brazilian Scientific Mobility Program Visiting Undergraduate Student); Trained in lab and conducted research on protein adsorption

96. **Mr. Ketley Alves** (Summer 2015; Brazilian Scientific Mobility Program Visiting Undergraduate Student): Trained in lab and conducted research on biomass pretreatment and its impact on cellulase digestibility

High School Students

1. **Ms. Kathleen Zheng** (July-Aug 2025/2026; ACS SEED Summer Program High School Intern; Project Title "*Enzyme discovery for Sargassum seaweed degradation*"; Lab Mentor: Rebecca Garcia)
2. **Ms. Bree Zavala** (July-Aug 2025; ACS SEED Summer Program High School Intern; Project Title "*Advanced manufacturing of protein based biological drugs*"; Lab Mentor: Aditya Narvekar)
3. **Mr. Aaron Romero** (July-Aug 2025; ACS SEED Summer Program High School Intern; Project Title "*Advanced manufacturing of carbohydrate-based prebiotics*"; Lab Mentor: Srividya Tallavajhula)
4. **Ms. Natasha Ramakrishnan** (July-Aug 2025; Rutgers Summer Program High School Intern; Project Title "*Advanced manufacturing of carbohydrate-based prebiotics*"; Lab Mentor: Srividya Tallavajhula)
5. **Ms. Genesis Martinez** (July-Aug 2024; ACS SEED Summer Program High School Intern"; Project Title "*Single-molecule characterization of glycan binding proteins*" Lab Mentor: Ryan Murphy)
6. **Ms. Nicole Onuoha** (July-Aug 2024; ACS SEED Summer Program High School Intern"; Project Title "*Advanced manufacturing of protein based biological drugs*"; Lab Mentor: Aditya Narvekar)

Doctoral Dissertation Committee Member at Rutgers

Current

1. Ms. Xinyi Li (June 2026; Rutgers CBE PhD Dissertation and Defense Committee Member "*Bioseparation of Enzymes in Synthetic Membraneless Compartments for Biocatalysis Studies*"; PhD Thesis Director - Dr. Benjamin Schuster)
2. Ms. Seyedeh Negin Hadisadeh (Fall 2028 Tentative Graduation; Rutgers CCB PhD Dissertation and Defense Committee Member "*TBD*"; PhD Dissertation Director - Dr. Sagar Khare)

Former

1. Mr. Lingjun Xie (Jan 2026; Rutgers CCB PhD Dissertation and Defense Committee Member "[*Control of enzymatic activity and stability*](#)"; PhD Dissertation Director - Dr. Sagar Khare)
2. Mr. Michael Xiaobin Shen (May 2025; Rutgers Food Science PhD Dissertation and Defense Committee Member "[*Interfacial dynamics and non-thermal processing: A multiscale approach to beverage formulation and nutrient delivery*](#)"; PhD Dissertation Director - Dr. Paul Takhistov)
3. Ms. Yuxin Liu (Oct 2022; Rutgers CBE PhD Dissertation and Defense Committee Member "[*Metabolic engineering of E. coli for biosynthesis of 4-hydroxybenzoate and its derivative hydroxyhydroquinone*](#)"; PhD Dissertation Director - Dr. Haoran Zhang)
4. Ms. Nanxia Zhao (May 2022; Rutgers CBE PhD Dissertation and Defense Committee Member "[*Design of amphiphilic macromolecule and antioxidant based nanotherapeutics for microglia-mediated neuroinflammation in synucleinopathy*](#)"; PhD Dissertation Director - Dr. Prabhas Moghe)
5. Ms. Ou Yang (May 2021; Rutgers CBE PhD Dissertation and Defense Committee Member "[*Process modeling and optimization of biopharmaceutical manufacturing*](#)"; PhD Dissertation Director - Dr. Marianthi Ierapetritou)
6. Mr. Shashwat Gupta (Sept 2020; Rutgers CBE PhD Proposal Exam Committee Member "[*Method development and application of high resolution vibrational spectroscopic techniques to elucidate*](#)

[the microstructure of pharmaceutical powder formulations](#)"; PhD Dissertation Director - Dr. Fernando Muzzio)

7. Mr. Abhay Athaley (Jan 2020; Rutgers CBE PhD Dissertation and Defense Committee Member "[Integrated design, analysis, and optimization of chemical production from biomass feedstocks](#)"; PhD Dissertation Director - Dr. Marianthi Ierapetritou)

Doctoral Dissertation Committee Member at Other Universities

1. Ms. Zeinab Amoabediny (June 2026; NJIT BME PhD Dissertation Defense Committee External Member "TBD"; PhD Thesis Director - Dr. Alexander Buffone Jr.)
2. PhD Dissertation Confidential External Evaluator for IIT Delhi School of Interdisciplinary Research (May 2025)
3. Mr. Youngwoo Woo (May 2021; University of Michigan ChemE PhD Thesis Proposal Title "[Computer-aided enzyme design for fucosylated oligosaccharides](#)"; PhD Thesis Director - Dr. Heather Mayes)

Students & Research Staff Supervised Previously at MSU and UW-Madison (GLBRC)

Graduate Students Co-Advised at Rutgers at MSU

1. **Ms. Carolyn Haarmeyer** (2012-2016; Michigan State CHEMS MS); Co-advised MS graduate student with Prof. Tim Whitehead (Primary Advisor) at MSU.
2. **Ms. Stephanie Crews** (2011-2013; Michigan State CHEMS MS); Co-advised MS graduate student with Prof. Bruce Dale (Primary Advisor) at MSU.

Research Associate Staff Supervised or Trained at MSU (GLBRC)

1. **Mr. James Humpula** (2009-2012; Michigan State CHEMS Research Associate)
2. **Ms. Margaret Magyar** (2010-2012; Michigan State CHEMS Research Associate)
3. **Mr. Nirmal Uppugundla** (2010-2012; Michigan State CHEMS Research Associate)
4. **Ms. Christa Gunawan** (2010-2012; Michigan State CHEMS Research Associate)
5. **Mr. Nana Achampong** (2007-2009; Michigan State CHEMS Research Associate)

High School & Undergraduate Students Supervised or Trained at UW-Madison (GLBRC)

1. **Ms. Amy (Dohi) Lim** (2012-2015; Wisconsin-Madison Biochemistry BS)
2. **Ms. Alice Zhang** (Summer 2015; High School Junior Student)
3. **Ms. Crystal Dyer** (Summer 2013; High School Junior Student)
4. **Mr. Alec Hernst** (2012-2014; Wisconsin-Madison Biochemistry BS)
5. **Ms. Cassandra Hainer** (2012-2014; Wisconsin-Madison Biochemistry BS)
6. **Mr. Dylan Edwards** (2012-2014; Wisconsin-Madison Biochemistry BS)
7. **Mr. Cameron Seiser** (2012-2014; Wisconsin-Madison Biochemistry BS)
8. **Mr. Sungsoo Lim** (2012-2014; Wisconsin-Madison Biochemistry BS)

High School Teacher Supervised or Trained at UW-Madison (GLBRC)

1. **Mr. Marin Dobson** (Summer 2013; High School Teacher; GLBRC RET Scholar)

High School & Undergraduate Students Supervised or Trained at MSU (GLBRC)

1. **Mr. Thomas Birkett** (2009-2011; Michigan State CHEMS BS)
2. **Mr. Chad Rogers** (2008-2010; Michigan State CHEMS BS)
3. **Mr. Aditya Rajagopalan** (Summer of 2008 and 2009; High School Junior/Senior Student)
4. **Mr. Luke Biela** (2007-2009; Michigan State CHEMS BS)
5. **Ms. Linpei Chang** (2007-2009; Michigan State CHEMS BS)
6. **Ms. Linda Stanek** (2006-2008; Michigan State CHEMS BS)

Undergraduate and Graduate Courses Taught at Rutgers

Course Details

[155:298](#) Professional Skills (Fall 2023)

[155:324](#) Separation Processes (Spring 2015-Present)

[155:507](#) Analytical Methods in Chemical & Biochemical Engineering (Fall 2016-2018)

[155:411](#) Biochemical Engineering: Co-instructor with Haoran Zhang (Fall 2020)

[155:532](#) Engineering of Sugars: Glycosciences in Health, Energy, and Materials (Fall 2021, 2024, 2026)

[155:601/602](#) Rutgers CBE Graduate Seminar Series (2020-Present)

Non-Rutgers personnel who do not have access to Sakai/Canvas can review most recent representative course syllabi on CBE undergraduate/graduate courses website or Chundawat Research Website here:

<https://chundawat.rutgers.edu/teaching/>

Other Course and Curricular Development at Rutgers

1. *CBE Process Engineering Laboratory Undergraduate Lab Course (155:416)*: In collaboration with Dr. Alex Bertuccio, successfully integrated a new biomass conversion experiment in process lab curriculum focused on cellulase kinetics modeling relevant to biochemical/bioprocess engineering.
2. *CBE Separation Processes Undergraduate Course (155:324)*:
 - a. In collaboration with Dr. Alex Bertuccio, successfully integrated an ethanol-water separation experimental project into 155:324 course in Spring 2019. Students work on a pilot-scale wiped film evaporator to collect data and model system using Aspen Plus software.
 - b. Programmatic integration of Learning Assistants (LA's), sponsored by the Rutgers Learning Centers, into separations process course in Spring 2015 to improve student learning and performance. LA's meet with instructor on a weekly basis to provide updates.

Invited or Guest Instructor at Rutgers

1. *Guest instructor for BME Graduate Course on Topics in Advanced Biotechnology I*: 3-hour seminar on 'Glycoengineering for a sweeter and healthier future' in Prof. Martin Yarmush's and Prof. Ann Stock's Rutgers Biotechnology Training Program course (16:125:604) (Spring 2023).
2. *Guest instructor for CBE Graduate Course on Biochemical Engineering*: 3-hour seminar on 'Enzyme/Protein Engineering' in Prof. Charlie Roth's Biochemical Engineering course (155:531) (Spring 2019, Fall 2020, Fall 2022).

3. *Guest instructor for CBE Undergraduate Course on Professional Skills Development:* Professional presentation skills 80 min seminar in Prof. Marianthi Ierapetritou's Professional Skills Development course (155:298) for Rutgers CBE sophomores (Fall 2015, 2016, 2017).
4. *Guest instructor for School of Engineering CBE Introductory and Outreach Courses:*
 - a. Lecture/demo activity (80 min) introducing chemical and biochemical engineering to Rutgers School of Engineering first year undergraduates (Fall 2016, 2017, 2018, 2023) as part of Introduction to Engineering Orientation Course (14:440:100).
 - b. Introduction to engineering lecture (80 min) for Rutgers women undergraduates at the Reilly Douglass Engineering Living-Learning Community (Fall 2016).

Undergraduate and Graduate Courses Taught at Michigan State University

CHE-891: Plant Cell Wall Chemistry 3-credit graduate elective course (Department of Chemical Engineering & Materials Science) on fundamental and applied aspects of the chemistry of plant cell walls and their conversion to fuels and chemicals co-instructed with David Hodge in Spring 2012.

CSE-101: Computing Concepts & Competencies 3-credit undergraduate core course (Department of Computer Science and Engineering) taught as sole TA instructor in Fall 2004 and Spring 2005.

Other K-12 Education and Outreach Instructional Materials Development

1. *Development of K-12 Glycoengineering For Gut Health Outreach Activity at Rutgers:* Developed a hands-on demonstration activity titled 'Unlocking the Energy Code: All about Carbohydrates & Enzymes' to introduce students to the role of Carbohydrate-Active enzymes (CAZymes) and prebiotics in human health. The activity features a student-developed enzyme assay using over-the-counter reagents to detect lactose and evaluate the presence of Human Milk Oligosaccharides (HMOs) in commercial beverages and infant formulas. This outreach program was launched July 9th 2024 for ~30 rising 9th-grade school students in collaboration with the Rutgers Academy for Girls in Engineering & Technology (TARGET) and the Project SUPER undergraduate research program. This activity, supported by the National Science Foundation (NSF) awards 1904890/1904862, helped translate fundamental research on HMO biosynthesis into educational modules designed to inspire interest in the burgeoning fields of glycosciences and glycoengineering. Detailed educational materials relevant to this student activity are available [online](#) and upon request.
2. *Development of Biophysics Outreach Activity at Rutgers:* Developed a MATLAB based program for visualization of tethered particle dynamics and single-molecule biophysics experimental data analysis. Detailed educational materials relevant to this student activity are available [online](#).
3. *Development of Pharmaceutical Engineering Outreach Activity at Rutgers:* Developed a hands-on demonstration activity for visualizing granular flow and fluidization of solids with relevance to API/tablet drug manufacturing. Hands-on 2-hr activity deployed in 2023 fall with over 850 first year SOE students. Educational materials for this event are available online.
4. *STEM Ambassador Outreach Activity at Rutgers:* Participated in 2-hr Zoom-based K-12 outreach activity on 29th July 2020 with over 40 high school students from local New Jersey schools as STEM ambassador and scientist. Details about this Rutgers NJAES sponsored annual summer STEM program are available upon request.

5. *Development of K-12 Biofuels and Enzyme Engineering Outreach Activity at Rutgers*: Developed a K-12 hands-on demonstration activity on using cellulase enzymes for biomass conversion. Hands-on 4-hr activity deployed in 2017 and 2018 summer with over 45 high school students from local New Jersey schools. Detailed educational materials relevant to this hands-on student activity are now available [online](#).
6. *Research Education for Teachers (RET) Mentor at GLBRC*: Research mentor for **Mr. Marin Dobson**, a biology and biotechnology teacher at Fort Atkinson high school (Wisconsin), as part of the Research Experience for Teachers (RET) program sponsored by the Department of Energy's Great Lakes Bioenergy Research Center (DOE-GLBRC) at the University of Wisconsin-Madison for seven weeks in the summer of 2013. All relevant educational material on biomass deconstruction is made available via [GLBRC](#).

Fellowships, Awards, and/or Honors Earned by Chundawat Research Group Members at Rutgers

Postdoctoral Scholars

1. Award Recipient: **Dr. Aditya Narvekar**, Sponsor: Bioprocess International Conference (Boston), Date: Sept 2025. Best Research Poster Award.
2. Fellowship Recipient: **Dr. Zhao Chao**, Post-doctoral Research Advisor: Shishir Chundawat (Rutgers University), Project Title: *Catalytic routes to upgrade glycan-derived substrates to value-added products*, Fellowship Amount: \$30,000, Start Date: Aug 2018, End Date: Jul 2019, Agency: China Scholarship Council from Government of China, Role: Post-doctoral Research Advisor
3. Fellowship Recipient: **Dr. Shyamal Roy**, Post-doctoral Research Advisor: Shishir Chundawat (Rutgers University), Project Title: *Integrated cellulosic biomass pretreatment and catalytic-upgrading processes for advanced biofuel production*, Fellowship Amount: \$37,200, Start Date: July 2016, End Date: June 2017, Agency: University Grant Commission (UGC) Raman Postdoctoral Fellowship from Government of India, Role: Post-doctoral Research Advisor
4. Fellowship Recipient: **Dr. Yongling Qin**, Post-doctoral Research Advisor: Shishir Chundawat (Rutgers University), Project Title: *Conversion of renewable biomass to fuels using a thermophilic cellulolytic microbe Thermobifida fusca*, Fellowship Amount: \$32,400, Start Date: Jan 2016, End Date: Jan 2017, Agency: Guangxi Scholarship from Guangxi Education Department from Government of China, Role: Post-doctoral Research Advisor

Graduate Students

1. Fellowship Recipient: **Mr. Austin Seamann**, Doctoral Research Advisor: Sagar Khare and Shishir Chundawat (Rutgers University), Annual Fellowship Amount: \$36,000, Start Date: Sept 2024, End Date: Aug 2026, Agency: NIH/Rutgers Biotechnology Training Grant, Role: Predoctoral Fellow
2. Award Recipient: **Mr. Dharanidaran Jayachandran**, Sponsor: Gordon Research Conference (GRC), Date: July 2023. Outstanding Research Award at the GRC Meeting on CAZymes for Glycan Conversions (Andover, NH).
3. Fellowship Recipient: **Mr. Antonio DeChellis**, Doctoral Research Advisor: Shishir Chundawat (Rutgers University), Annual Fellowship Amount: \$33,999, Start Date: Sept 2022, End Date: Aug 2024, Agency: NIH/Rutgers Biotechnology Training Grant, Role: Predoctoral Fellow
4. Award Recipient: **Mr. Dharanidaran Jayachandran**, Sponsor: Rutgers BME Annual Research Symposium, Date: Dec 2022. Outstanding poster award.

5. Award Recipient: **Mr. Aron Gyorgypal**, Sponsor: BioPro World Talent Campus, Date: Sept 2022. BioPro World Talent Meeting Participant/Fellow.
6. Award Recipient: **Mr. Aron Gyorgypal**, Sponsor: ORISE, Date: May-Sept 2022. ORISE Research Summer Intern at FDA CDER Office of Biotechnology Products.
7. Award Recipient: **Mr. Markus Hackl**, Doctoral Research Advisor: Shishir Chundawat (Rutgers University), NSF Internship Award, Spring 2021, Agency: NSF, Role: Internship at LUMICKS (remote)
8. Award Recipient: **Mr. Aron Gyorgypal**, Sponsor: International Foundation Process Analytical Chemistry Annual Meeting, Date: Feb 2020. Outstanding poster award.
9. Award Recipient: **Mr. Aron Gyorgypal**, Sponsor: Rutgers BME Annual Research Symposium, Date: Dec 2019. Outstanding poster award.
10. Award Recipient: **Mr. Bhargava Nemmaru**, Doctoral Research Advisor: Shishir Chundawat (Rutgers University), NSF Internship Award, Spring 2020, Agency: NSF, Role: Internship at NREL (in-person)
11. Fellowship Recipient: **Mr. Satvik Sharma**, Sponsor: NASA New Jersey Space Grant Consortium Graduate Student Fellowship, Start Date: Sept 2015, End Date: May 2016. This fellowship provided support for his annual stipend /year) at Rutgers.

Undergraduate and High School Students

1. Award Recipient: **Ms. Sofia Colella**, Research Advisor: Shishir Chundawat (Rutgers University), Fellowship Amount: \$3,000, Start Date: May 2025, End Date: Aug 2025, Agency: Rutgers Douglass WiSE SUPER Program, Role: SUPER Intern Research Advisor
2. Award Recipient: **Ms. Merlyn Roy**, Research Advisor: Shishir Chundawat (Rutgers University), Fellowship Amount: \$3,000, Start Date: May 2025, End Date: Aug 2025, Agency: Rutgers Douglass WiSE SUPER Program, Role: SUPER Intern Research Advisor
3. Award Recipient: **Ms. Raadha Garg**, Research Advisor: Shishir Chundawat (Rutgers University), Fellowship Amount: \$3,000, Start Date: May 2025, End Date: Aug 2025, Agency: Rutgers Douglass WiSE SUPER Program, Role: SUPER Intern Research Advisor
4. Internship Recipient: **Mr. Axel Yvoz**, Research Advisor: Shishir Chundawat (Rutgers University), Project Title: *Expression Purification and Characterization of Carbohydrate Binding modules by Acoustic Force Spectroscopy*, Start Date: May 2025, End Date: Aug 2025, Agency: Big Ten Academic Alliance (BTAA) International Research Experience Program (IREP), Role: BTAA-IREP Program Research Advisor
5. Internship/Award Recipient: **Ms. Genesis Martinez**, Research Advisor: Shishir Chundawat (Rutgers University), Project Title: *Protein Characterization Using Single Molecule Force Spectroscopy*, Start Date: July 2024, End Date: Aug 2024, Agency: American Chemical Society (ACS) SEED Summer Program, Role: High School Intern Advisor
6. Internship/Award Recipient: **Ms. Nicole Onuoha**, Research Advisor: Shishir Chundawat (Rutgers University), Project Title: *Glycobiologics Manufacturing and Characterization*, Start Date: July 2024, End Date: Aug 2024, Agency: American Chemical Society (ACS) SEED Summer Program, Role: High School Intern Advisor
7. Internship/Award Recipient: **Ms. Nurulhuda Binte Suaini**, Research Advisor: Shishir Chundawat (Rutgers University), Project Title: *PAT for Advanced GlycoBiologics Manufacturing*, Start Date: Sept 2024, End Date: Aug 2025, Agency: Turing Foundation and University of Surrey (UK), Role: Intern Research Advisor

8. Award Recipient: **Ms. Arushi Biswas**, Research Advisor: Shishir Chundawat (Rutgers University), Fellowship Amount: \$3,000, Start Date: May 2024, End Date: Aug 2024, Agency: Rutgers Douglass WiSE SUPER Program, Role: SUPER Intern Research Advisor
9. Award Recipient: **Ms. Iris Brody**, Research Advisor: Shishir Chundawat (Rutgers University), Fellowship Amount: \$3,000, Start Date: May 2024, End Date: Aug 2024, Agency: Rutgers Douglass WiSE SUPER Program, Role: SUPER Intern Research Advisor
10. Award Recipient: **Ms. Mahati Salapu**, Research Advisor: Shishir Chundawat (Rutgers University), Fellowship Amount: \$3,000, Start Date: May 2024, End Date: Aug 2024, Agency: Rutgers Douglass WiSE SUPER Program, Role: SUPER Intern Research Advisor
11. Award Recipient: **Ms. Sophia Fernandez**, Research Advisor: Shishir Chundawat (Rutgers University), Fellowship Amount: \$3,000, Start Date: May 2024, End Date: Aug 2024, Agency: Rutgers Douglass WiSE SUPER Program, Role: SUPER Intern Research Advisor
12. Internship Recipient: **Ms. Annouck Bonneau-Arene**, Research Advisor: Shishir Chundawat (Rutgers University), Project Title: *Enzyme Engineering for Human Milk Oligosaccharides Synthesis*, Start Date: May 2024, End Date: Aug 2024, Agency: Big Ten Academic Alliance (BTAA) International Research Experience Program (IREP), Role: BTAA-IREP Program Research Advisor
13. Internship Recipient: **Ms. Tiphaine Alves Cunha**, Research Advisor: Shishir Chundawat (Rutgers University), Project Title: *Enzyme Engineering for Waste Biomass and Plastics Upcycling*, Start Date: June 2024, End Date: Aug 2024, Agency: Big Ten Academic Alliance (BTAA) International Research Experience Program (IREP), Role: BTAA-IREP Program Research Advisor
14. Award Recipient: **Ms. Nabanita Paul** (Visiting Undergraduate Intern from IIT-Mumbai), Sponsor: Khorana Program Research Scholar/Intern Award, Summer 2024.
15. Fellowship Recipient: **Ms. Nakeitha Brown**, Research Advisor: Shishir Chundawat (Rutgers University), Project Title: *Advanced Analytical Analysis of mAb N-Glycans*, Fellowship Amount: \$25,000, Start Date: Aug 2023, End Date: Aug 2024, Agency: Post-Baccalaureate Research Experience for LSAMP Students (PRELS) Program at Rutgers-Newark, Role: PRELS Research Advisor
16. Award Recipient: **Mr. Gokulnath Ganesan** (Visiting Undergraduate Intern from IIT-Madras), Sponsor: Khorana Program Research Scholar/Intern Award, Summer 2023.
17. Award Recipient: **Ms. Saranya Saini Guttikonda**, Research Advisor: Shishir Chundawat (Rutgers University), Fellowship Amount: \$3,000, Start Date: May 2023, End Date: Aug 2023, Agency: Rutgers Douglass WiSE SUPER Program, Role: SUPER Intern Research Advisor
18. Award Recipient: **Ms. Olivia Reste**, Research Advisor: Shishir Chundawat (Rutgers University), Fellowship Amount: \$3,000, Start Date: May 2022, End Date: Aug 2022, Agency: Rutgers Douglass WiSE SUPER Program, Role: SUPER Intern Research Advisor
19. Award Recipient: **Ms. Jenna Douglass**, Sponsor: AIChE 2021 Annual Meeting Undergraduate Research Poster Competition (2nd Best Student Poster), Oct 2021.
20. Fellowship Recipient: **Ms. Shoili Banerjee**, Sponsor: Rutgers Energy Institute (REI) Summer Internship, Start Date: May 2020, End Date: Aug 2021.
21. Award Recipient: **Mr. Varun Raghuraman**, Sponsor: Aresty Undergraduate Research Fellowship, Start Date: Sept 2019, End Date: May 2020.
22. Fellowship Recipient: **Mr. Allan Wang**, Sponsor: Rutgers Energy Institute (REI) Summer Internship, Start Date: May 2018, End Date: Aug 2018.
23. Award Recipient: **Mr. Patrick Doran**, Sponsor: Aresty Undergraduate Research Fellowship, Start Date: Sept 2017, End Date: May 2018.

24. Fellowship Recipient: **Mr. Varun Raghuraman**, Sponsor: Aresty Undergraduate Summer Science Fellowship, Start Date: May 2017, End Date: Aug 2017.
25. Award Recipient: **Ms. Marissa Berger**, Sponsor: Aresty Summer Science and Undergraduate Research Fellowship, Start Date: Sept 2016, End Date: May 2017.
26. Award Recipient: **Ms. Cindy Farino**, Sponsor: Aresty Undergraduate Research Fellowship, Start Date: Sept 2016, End Date: May 2017.
27. Fellowship Recipient: **Ms. Alina Thokaddam**, Sponsor: Aresty Undergraduate Summer Science Fellowship, Start Date: May 2016, End Date: Aug 2016.
28. Fellowship Recipient: **Ms. Natália Dias**, Sponsor: Brazilian Scientific Mobility Program for Visiting Undergraduate Students, Start Date: May 2016, End Date: Aug 2016.
29. Fellowship Recipient: **Ms. Samantha Cobos**, Sponsor: NSF-REU and Rutgers RiSE Fellow Program, Start Date: May 2016, End Date: Aug 2016.
30. Fellowship Recipient: **Ms. Marissa Berger**, Sponsor: Aresty Undergraduate Summer Science Fellowship, Start Date: May 2015, End Date: Aug 2015.
31. Fellowship Recipient: **Mr. Ketley Alves**, Sponsor: Brazilian Scientific Mobility Program for Visiting Undergraduate Students, Start Date: May 2015, End Date: Aug 2015.

Conference Presentations, Lectures, and Interviews

Invited Plenary Talks & Seminars

1. Invited seminar by **Shishir Chundawat** at Society for Industrial Microbiology and Biotechnology (SIMB) Symposium on Biomaterials, Fuels and Chemicals (SBFC) Annual Meeting. *Harnessing the 'Sweet' Rules of Life: Carbohydrate-Active Enzyme Design and Engineering at Automated BioFoundries for Waste Upcycling*, New Orleans, May 2025. (*presented by Rebecca Garcia)
2. Invited seminar by **Shishir Chundawat** at Rutgers State University of New Jersey (Rutgers Tri-departmental-Pharmacology, Biochemistry & Molecular Biology, and Neuroscience & Cell Biology-Seminar), *Carbohydrate-Active Enzyme (CAZyme) Engineering for Biomanufacturing of Designer Foods, Fuels, and Biologics*, New Brunswick, Mar 2026.
3. Invited seminar by **Shishir Chundawat** at Rutgers State University of New Jersey (Food Science Department), *Carbohydrate-Active Enzyme (CAZyme) Engineering for Biomanufacturing of Designer Foods, Fuels, and Biologics*, New Brunswick, Dec 2025.
4. Invited seminar by **Shishir Chundawat** at City College of New York (Dept. of Chemical Engineering), *Carbohydrate-Active Enzyme (CAZyme) Engineering for Biomanufacturing of Designer Foods, Fuels, and Biologics*, New York City, Dec 2025.
5. Invited plenary talk by **Shishir Chundawat** at Gordon Research Conference on Carbohydrate-Active Enzymes for Glycan Conversions, *CAZyme Engineering Guided by Computational Protein Design, Automated Biofoundry Screening, and Single-Molecule Assays*, Andover (NH), July 2025.
6. Invited talk by **Shishir Chundawat** at the National Institute for Innovation in Manufacturing Biopharmaceuticals (NIIMBL) Annual Meeting, *PC5.2-112 - An Integrated Platform for Fully Automated, Continuous, & Real-Time Multi-Attributes Monitoring of Upstream Processes for Glycosylated Monoclonal Antibodies Production*, Washington DC (DC), June 2025.

7. Invited talk by **Shishir Chundawat** at Rutgers State University of New Jersey (RCEI sponsored conference on Advances in Research on Seaweeds in the Caribbean), *Seaweed waste-to-bioproducts guided by artificial intelligence and synthetic biology*, Piscataway, May 2025.
8. Invited talk by **Shishir Chundawat** at American Chemical Society (ACS) Annual Middle Atlantic Region Meeting (MARM) session “Unlocking Glycosylation”, *Sugar sugar everywhere...but not a spoonful for my drink*, Seton Hall (NJ), May 2025.
9. Invited seminar by **Shishir Chundawat** at Rutgers State University of New Jersey (Institute of Quantitative Biomedicine), *Sugar sugar everywhere...but not a spoonful for my drink: Engineering a sweeter, healthier, & cleaner future*, Piscataway, Nov 2024.
10. Invited seminar by **Shishir Chundawat** at New Jersey Institute of Technology (Dept. of Biomedical Engineering), *Sugar sugar everywhere...but not a spoonful for my drink*, Newark, Nov 2024.
11. Invited talk by **Shishir Chundawat** at the National Institute for Innovation in Manufacturing Biopharmaceuticals (NIIMBL) Annual Meeting, *PC5.2-112 - An Integrated Platform for Fully Automated, Continuous, & Real-Time Multi-Attributes Monitoring of Upstream Processes for Glycosylated Monoclonal Antibodies Production*, Washington DC (DC), June 2024.
12. Invited webinar by **Shishir Chundawat** at 6th Annual Event in the Bioprocessing Virtual Event Series (LabRoots), *An automated and fully integrated process analytical platform for combined analysis of N-glycans and amino acids to enable continuous manufacturing of glycosylated biologics*, Virtual Event, April 2024 (Jointly presented with Aditya Narvekar).
13. Invited symposium select talk by **Shishir Chundawat** at Biophysical Society Annual Meeting, *Visualizing Cellulose Polysaccharides Biosynthesis and Dynamic Assembly Into Cell Walls During Live Plant Cell Imaging*, Philadelphia (PA), 2024.
14. Invited plenary talk by **Shishir Chundawat** at Gordon Research Conference on Carbohydrate-Active Enzymes for Glycan Conversions, *Mr. Cell, Tear Down This Wall: Visualizing Cellulose Biosynthesis, Assembly, & Degradation*, Andover (NH), 2023.
15. Invited webinar by **Shishir Chundawat** at 5th Annual Event in the Bioprocessing Virtual Event Series (LabRoots), *N-GLYcanalyzer: An Automated Process Analytical Platform To Enable Continuous Manufacturing Of Biologics*, Virtual Event, April 2023 (Jointly presented with Aron Gyorgypal).
16. Invited seminar by **Shishir Chundawat** at University of Rhode Island (*Department of Chemistry*), *Glycoengineering for a cleaner, healthier, & sweeter future*, Jan 2023 (Zoom Webinar Presented Remotely On-line).
17. Invited seminar by **Shishir Chundawat** at 22nd Annual PepTalk Meeting session “Higher-Throughput Bioproduction”, *Integrated Process Analytical Platform for Automated Monitoring of Monoclonal Antibody N-Glycosylation*, San Diego, Jan 2023 (Presented by Student Aron Gyorgypal).
18. Invited seminar by **Shishir Chundawat** at Indian Institute of Technology-Madras (*Department of Biotechnology, Bhupat & Jyoti Mehta School of Biosciences*), *Glyco-engineering for a cleaner, sweeter, and healthier future!*, Jan 2023 (Zoom Webinar Presented Remotely On-line).
19. Invited seminar by **Shishir Chundawat** at Michigan Technological University (Dept. of Chemical Engineering), *Glycoengineering for a cleaner, healthier, & sweeter future*, Dec 2022 (Zoom Webinar Presented Remotely On-line).
20. Invited seminar by **Shishir Chundawat** at University of Delaware (Dept. of Chemical & Biomolecular Engineering), *Glycoengineering for a cleaner, sweeter, & healthier future*, Newark, Nov 2022.

21. Invited webinar by **Shishir Chundawat** at the 6th Annual United States Pharmacopeia (USP) Workshop on Biologics and Peptides, *PAT Toolkit for Monitoring of Monoclonal Antibody (mAb) N-Glycosylation in Real-Time*, Online, June 2022 (Zoom Summit Held Remotely On-line).
22. Invited webinar by **Shishir Chundawat** at Oak Ridge National Laboratory, *Visualizing plant biomass and renewable plastics biosynthesis and deconstruction to enable advances in Biological and Environmental Science research at STS*, Oak Ridge, June 2022.
23. Invited seminar by **Shishir Chundawat** at Indian Institute of Technology (Dept. of Chemical Engineering), *Next generation glycoengineering for a cleaner, healthier, & sweeter future for humanity*, New Delhi, June 2022.
24. Invited plenary seminar by **Shishir Chundawat** at the 44th Symposium on Biotechnology for Fuels and Chemicals in plenary session on “Biomass-active enzyme discovery, mechanisms, and engineering”, *Watching slow-crawler cellulolytic enzymes to enable giant leaps for biomass deconstruction*, New Orleans, May 2022.
25. Invited webinar by **Shishir Chundawat** for Process Analytical Technology (PAT) community of American Association of Pharmaceutical Scientists (AAPS), *An Integrated Process Analytical Platform for Automated Monitoring of Monoclonal Antibody N-linked Glycosylation*, Online, February 2022 (Zoom Summit Held Remotely On-line).
26. Invited seminar/webinar by **Shishir Chundawat** for Process Analytical Technology (PAT) community at the WCBP 2022 Annual Meeting (The 26th Symposium on the Interface of Regulatory and Analytical Sciences for Biotechnology Health Products), *N-GLYcanalyzer: PAT Tool for Near Real-Time Monitoring of Monoclonal Antibody (mAb) Glycosylation*, Washington DC, January 2022 (Hybrid Summit Held In-Person and Remotely On-line).
27. Invited seminar by **Shishir Chundawat** at 21st Annual PepTalk Meeting session “Cell Line Engineering & Development”, *Engineered Regulon to Enable Autonomous Azide Ion Biosensing, Recombinant Protein Production, and in Vivo Glycoengineering*, San Diego, Jan 2022 (Hybrid Summit Held In-Person and Remotely On-line).
28. Invited seminar by **Shishir Chundawat** at Gordon Research Conference on Carbohydrate-Active Enzymes for Glycan Conversions, *Engineering and mechanistic analysis of CAZymes for in-vitro and in-vivo synthesis of glycan polymers*, Andover, July 2021 (Conference Postponed to July 2023 due to SARS-CoV2 pandemic).
29. Invited seminar at Department of Energy (DOE) 2021 Bioimaging Science Program Annual Principal Investigator (PI) Meeting, *In planta multimodal single-molecule imaging of cell wall synthesis*, Virtual Meeting, Feb 2021. Attended by PI Sang-Hyuk Lee & Co-PI Chundawat.
30. Invited seminar by **Shishir Chundawat** at Washington University at St. Louis (Dept. of Energy, Environmental & Chemical Engineering), *Glyco-enzymes engineering for a sweeter, cleaner, and healthier future!*, Zoom Webinar, Dec 2020.
31. Invited seminar by **Shishir Chundawat** as part of the Global Colgate-Palmolive Tech Exchange Program, *Glyco-Engineering for a Sweeter & Healthier Future*, Piscataway, Oct 2020 (Webinar Held Remotely On-line Due to COVID-19 Pandemic).
32. Invited seminar by **Shishir Chundawat** at Agilent Biopharmaceutical Web Summit 2020 on Discovery Development and Characterization of Next-Generation Therapeutics, *Continuous Biomanufacturing at Rutgers University: Integrated Real-time PAT, Modeling, and Process Control Strategies to Enable Production of Glycosylated Biologics*, Piscataway, March 2020 (Summit Held Remotely On-line Due to COVID-19 Pandemic).

33. Invited seminar at Department of Energy (DOE) 2020 Bioimaging Science Program Annual Principal Investigator (PI) Meeting, *In planta single-molecule imaging and holographic force spectroscopy to study real-time, multimodal turnover dynamics of polysaccharides and associated carbohydrate metabolites*, Washington DC, Feb 2020. Attended by PI Sang-Hyuk Lee & Co-PI Chundawat.
34. Invited seminar at Food and Drug Administration (FDA) Center for Biologics Evaluation and Research (CBER) Program Advanced Technologies Symposium, *Advanced Continuous Upstream Manufacturing of Biotherapeutics*, Silver Spring, July 2019. Talk jointly presented by PI Marianthi Ierapetritou and Co-PI **Shishir Chundawat**.
35. Invited seminar by **Shishir Chundawat** at Gordon Research Seminar on Carbohydrates (Development of Carbohydrate-Based Therapeutics and Targets), *Engineering and Mechanistic Analysis of Novel Multidomain Transglycosylases*, Hong Kong, June 2019.
36. Invited seminar at Department of Energy (DOE) 2019 Bioimaging Science Program Annual Principal Investigator (PI) Meeting, *In planta single-molecule imaging and holographic force spectroscopy to study real-time, multimodal turnover dynamics of polysaccharides and associated carbohydrate metabolites*, Vienna, Feb 2019. Attended by PI Sang-Hyuk Lee & Co-PI Chundawat.
37. Invited seminar by **Shishir Chundawat** at New York Institute of Technology (NYIT) 13th Annual Energy Conference on Bioenergy and Natural Systems, *Engineering Enzymes to Reduce Cellulosic Biofuels Production Costs*, Old Westbury (New York), June 2018.
38. Invited fermentation series seminar by **Shishir Chundawat** at Rutgers State University of New Jersey (Dept. of Biochemistry and Microbiology), *Engineering 'small' enzyme steps to make giant leaps save our planet!*, New Brunswick, Nov 2017.
39. Invited seminar by **Shishir Chundawat** at Exxon-Mobil Research Division, *Deconstruction of cellulosic biomass to fuels and chemicals using an integrated approach*, Annandale (New Jersey), March 2017.
40. Invited seminar by **Shishir Chundawat** at Montclair State University, *Conversion of lignocellulosics to biofuels using a multistep chemo-enzymatic deconstruction approach*, Montclair (New Jersey), April 2017.
41. Invited poster presentation by **Shishir Chundawat** at Gordon Research Conference on Cellulases & Other Carbohydrate-Active Enzymes, *Single-molecule characterization of processive cellulase activity on cellulose*, Andover (NH), July 2017.
42. Invited seminar by **Shishir Chundawat** at Rutgers State University of New Jersey (Rutgers Global Grant Symposium), *Enabling the Creation of a Sustainable and Economically Viable Cellulosic Biofuels Industry in Brazil*, New Brunswick, Nov 2017.
43. Invited US Naval Research Laboratory (NRL) colloquium series seminar by **Shishir Chundawat** at US Naval Research Center (Center for Bio/Molecular Science and Engineering), *Providing a molecular basis to modeling protein adsorption to multimodal carbohydrate ligands*, Washington DC, Sept 2016.
44. Invited seminar by **Shishir Chundawat** at Rutgers State University of New Jersey (Dept. of Chemical & Biochemical Engineering), *Deconstruction of cellulosic biomass to fuels and chemicals: The emerging paradigm of substrate engineering*, New Brunswick, Mar 2014
45. Invited seminar by **Shishir Chundawat** at the 36th Symposium on Biotechnology for Fuels and Chemicals in plenary session on "Enzyme Science and Technology II – Modeling and Structure/Function Relationship", *Cellulases have low affinity but improved activity on unnatural cellulose allomorphs*, Clearwater Beach, May 2014.

46. Invited seminar by **Shishir Chundawat** at Iowa State University (Dept. of Chemical & Biological Engineering), *Deconstruction of cellulosic biomass to fuels and chemicals: The emerging paradigm of substrate engineering*, Ames, Feb 2014.
47. Invited seminar by **Shishir Chundawat** at American Institute of Chemical Engineers (AIChE) Annual Meeting session "Advances In Biofuels: DOE Bioenergy Research Centers", *Interactions of cellulolytic proteins with a non-native, polysaccharide allomorph formed during pretreatment of lignocellulosic biomass*, San Francisco, Nov 2013.
48. Invited seminar by **Shishir Chundawat** at American Crystallographic Association (ACA) Annual Meeting session "Materials for a Sustainable Future", *Modifying native crystalline polysaccharide ultrastructure can improve its chemical and biological processability to fuels, chemicals and materials*, Boston, July 2012.
49. Invited seminar by **Shishir Chundawat** at American Institute of Chemical Engineers (AIChE) Annual Meeting session "Advances In Biofuels: DOE Bioenergy Research Centers", *Development of the Extractive-AFEX (E-AFEX) pretreatment process*, Pittsburgh, Oct 2012.
50. Invited plenary talk by **Shishir Chundawat** at Materials Research Society (MRS) Spring Meeting (Renewable Fuels and Nanotechnology Session), *Impact of Ammonia Treatment on Natural Lignocellulosic Composites*, San Francisco, April 2011.
51. Invited plenary talk by **Shishir Chundawat** at Gordon-Kenan Research Seminar on Cellulosomes, Cellulases & Other Carbohydrate Modifying Enzymes, *Role of cellulose crystal structure on exo- and endo-cellulase synergistic activity*, Easton (MA), 2011.
52. Invited seminar by **Shishir Chundawat** at Institute of Chemical Technology (Dept. of Chemical Engineering), *Deconstruction of lignocellulosic biomass to fuels and chemicals*, Mumbai, Aug 2011.
53. Invited seminar by **Shishir Chundawat** at Michigan State University (Dept. of Chemical Engineering & Materials Science), *A new strategy for biomass deconstruction to fuels and chemicals*, East Lansing, Sept 2011.
54. Invited seminar by **Shishir Chundawat** at American Institute of Chemical Engineers (AIChE) Annual Meeting session "Advances In Biofuels: DOE Bioenergy Research Centers", *The binding properties of cellulases on cellulose allomorphs and pretreated biomass during enzymatic hydrolysis*, Minneapolis, Oct 2011.
55. Invited seminar by **Shishir Chundawat** at Department of Energy Bioenergy Science Center (BESC) Biomass Characterization Workshop, *Characterizing the effect of alkaline based pretreatments on lignocellulosic cell walls*, Riverside, Jan 2010.
56. Invited seminar by **Shishir Chundawat** at Los Alamos National Laboratory, *"NH₃"...a potent catalyst for deconstructing plant cell walls to fuels and chemicals*, Los Alamos, Sept 2010.
57. Invited seminar by **Shishir Chundawat** at Department of Energy Inter-Bioenergy Research Center Workshop (Enzymes for biomass degradation), *Effect of ammonia treatments on enzyme-biomass interactions*, Madison, Nov 2010.
58. Invited seminar by **Shishir Chundawat** at the DOE Great Lakes Bioenergy Research Center (GLBRC) Annual Retreat, *Assessing the ability of glycosyl hydrolases to digest ammonia pretreated biomass*, South Bend, May 2009.
59. Invited seminar by **Shishir Chundawat** at Verenium Corporation, *Towards a fundamental understanding of ammonia based pretreatment and enzymatic hydrolysis of lignocellulosic biomass*, San Diego, Feb 2008.

Contributed Talks (asterisk* indicates seminar talk presenter)

1. Choi D*, Garcia R, Choi S, Lee W, Jadhao PR, Vasquez RM, **Chundawat SPS**, Park AH, Moment A. *Sustainable Algal-Mining for the Recovery of Rare Earth Elements from Sargassum Seaweed*. AIChE Annual Meeting 2026, Minneapolis. (Pending Oral Presentation by Postdoc).
2. Anastasi A*, Masuku C, Ravikumar PK, **Chundawat SPS**, Hildebrandt D. *Estimation of Thermodynamic Properties for Cellulosic Biomass-Derived Compounds: Application to Heat and Work Balances in Process Simulation*. ESCAPE 36 - European Symposium on Computer Aided Process Engineering, June 2025, Sheffield (UK). (Pending Oral Presentation by Student).
3. Ravikumar PK*, Anastasi A, Masuku C, **Chundawat SPS**, Hildebrandt D. *Thermodynamic Attainable Region Framework for CO₂ Utilization and Energy Integration*. 30th Annual ACS Green Chemistry and Engineering Conference, June 2025, San Antonio (TX). (Pending Oral Presentation by Student).
4. Ali A, Nandi S, Choi D, Resheidat M, Echauri SG, Stephens T, Garcia R, Sanchez M, Roberson L, **Chundawat SPS**, Lee W, Choi S, Moment A, Bhattacharya D, Avalos J*. *Advances in Sargassum-associated microorganisms and rare earth element recovery toward a Sargassum biorefinery (SaBRe)*. Society for Industrial Microbiology and Biotechnology (SIMB) Symposium on Biomaterials, Fuels and Chemicals (SBFC) Annual Meeting 2025, May 2025, New Orleans. (Oral Presentation by Avalos).
5. Garcia R*, **Chundawat SPS**. *Taming the Tide: Transforming Sargassum Seaweeds into High Value Products*. Society for Industrial Microbiology and Biotechnology (SIMB) Symposium on Biomaterials, Fuels and Chemicals (SBFC) Annual Meeting 2025, May 2025, New Orleans. (Oral Presentation by Student).
6. **Chundawat SPS***. *Sargassum Seaweed Deconstruction: Updates from Year 2*. Sargassum BioRefinery (SaBRe) Center Annual Meeting 2026, Piscataway. (Oral Presentation by Chundawat in April 2026).
7. Narvekar A*, **Chundawat SPS**. *Glycosylated monoclonal antibody critical quality attributes determination using fully automated sample preparation and analytical workflows*. bioprocess International Annual Meeting 2025, Boston. (Oral Presentation by Narvekar in Sept 2025).
8. Avalos J*, Bhattacharya D, **Chundawat SPS**, Khare S, Kite-Powell H, Louime C, McGonagill A, Moment A, Park A, Roberson L, Ramírez DR, Yoshikuni Y. *SaBRe: A Sargassum BioRefinery to valorize uncontrolled blooms of brown macroalga*. International Seaweed Symposia (ISS) 2025 Meeting, Victoria (Canada). (Oral Presentation by Avalos in May 2025).
9. **Chundawat SPS***. *Sargassum Seaweed Deconstruction: Updates from Year 1*. Sargassum BioRefinery (SaBRe) Center Annual Meeting 2025, Princeton. (Oral Presentation by Chundawat in Jan 2025).
10. Reddy JV*, Leibiger T, Singh SK, Gyorgypal A, Narvekar A, **Chundawat SPS**, Papoutsakis ET, Ierapetritou M. *Investigating the effect of critical process parameters and media additives on site specific N-linked glycosylation of monoclonal antibodies*. AIChE Annual Meeting 2024, San Diego. (Oral Presentation by Student Jayanth Reddy in Oct 2024).
11. Kumar M*, **Chundawat SPS**. *Glycosynthase Engineering for Fucosylated Oligosaccharides Synthesis*. Rutgers BME BESS Annual Meeting 2022, Piscataway. (Oral Presentation by Student Mohit Kumar)
12. Reddy JV*, Raudenbush K, Yang O, Gyorgypal A, Chaturvedi A, **Chundawat SPS**, Ierapetritou M. *Model based insights on the effect of bioreactor pH and Temperature on N-Linked glycosylation of*

- mAbs produced by CHO cells.* AICHE Annual Meeting 2022, Phoenix. (Oral Presentation by Student Jayanth Reddy in Nov 2022).
13. **Chundawat SPS***, Bandi CK, Kumar M. *Engineered CynR Regulon for Enzymatic Biocatalysis Enables Milk Oligosaccharides Biosynthesis Using Azido Sugars.* AICHE Annual Meeting 2022, Phoenix. (Oral Presentation by Chundawat in Nov 2022).
 14. **Chundawat SPS***, Gyorgypal A. *N-GLYcanalyzer: An Integrated At-Line Process Analytical Toolkit for Enabling Continuous Biologics Manufacturing.* AICHE Annual Meeting 2022, Phoenix. (Oral Presentation by Chundawat in Nov 2022).
 15. Douglass J*, Nemmaru B, Shankar S, **Chundawat SPS.** *Rational Engineering of Cellulase Enzymes for Improved Biomass Conversion Via Reduced Non-Productive Binding.* AICHE Annual Meeting 2021, Boston. (Presentation by Student In-Person in Oct 2021)
 16. Yang O*, Reddy JV, Raudenbush K, Gyorgypal A, **Chundawat SPS**, Ierapetritou M. *Model-based Investigation of Upstream CHO Cell Culture Process for Production of Monoclonal Antibodies with Desired N-linked Glycosylation.* AICHE Annual Meeting 2021, Boston, In-person Meeting. (Presentation by Student in Oct 2021)
 17. Gyorgypal A*, **Chundawat SPS.** *A sequential injection based on-line PAT for real-time glycosylation monitoring of monoclonal antibodies.* AICHE Annual Meeting 2021, Boston, Virtual Meeting. (Online Presentation by Student in Oct 2021).
 18. Nemmaru B*, Douglass J, Yarbrough JM, Johnson MM, Lang MM, **Chundawat SPS.** *Rational engineering of polysaccharide-degrading enzymes: A case study on cellulases for improved biomass conversion to biofuels.* ACS Annual Meeting 2021, San Diego. (Online Student Presentation in Aug 2021)
 19. Nemmaru B*, **Chundawat SPS.** *Rational Engineering of Polysaccharide-Degrading Enzymes: A Case Study on Cellulases.* PEGS Annual Meeting 2021, San Diego. (Online Student Presentation in May 2021)
 20. Bandi CK*, **Chundawat SPS.** *Development of High-Throughput Cell Screening Toolkit for the Directed Evolution of Glycosynthase Enzymes for Bespoke Oligosaccharides Synthesis.* AICHE Annual Meeting 2020, Virtual Meeting. (Online Presentation by Student in Nov 2020)
 21. Hackl M*, Nemmaru B, **Chundawat SPS.** *Single-molecule protein-carbohydrate dissociation characterization using acoustic force spectroscopy.* ACS Annual Meeting 2020, Philadelphia. (Oral Presentation Cancelled due to COVID-19)
 22. Chopda V*, Gyorgypal A, Said R, Alter A, **Chundawat SPS**, Zhang H. *Continuous monoclonal antibody (mAb) biomanufacturing and integrated process analytics for mAb N-linked glycans analysis.* ACS Annual Meeting 2020, Philadelphia. (Oral Presentation Cancelled due to COVID-19)
 23. Bandi CK*, Burgin T, Pasture M, Goncalves A, Pingali SV, Gao J, Mayes H, **Chundawat SPS.** *Engineering and mechanistic analysis of carbohydrate binding domain assisted transglycosylation reactions for a novel protein design of multidomain transglycosidases.* ACS Annual Meeting 2020, Philadelphia. (Oral Presentation Cancelled due to COVID-19)
 24. Bandi CK*, Agrawal A, **Chundawat SPS.** *Development of a high-throughput cell screening toolkit for the directed evolution of glycosynthase enzymes for bespoke oligosaccharides synthesis using azido sugars.* ACS Annual Meeting 2020, Philadelphia. (Oral Presentation Cancelled due to COVID-19)
 25. Chopda V*, Gyorgypal A, Said R, Alter A, Zhang H, **Chundawat SPS.** *Optimizing monoclonal antibody (mAb) production: Establishing an in-house platform for continuous biomanufacturing and*

- integrated process analytics. Rutgers University (RU) BME Department Graduate Research Symposium, Dec 2019, Piscataway. (Oral Presentation by Staff)
26. Agrawal A*, **Chundawat SPS**. Development of click chemistry based screening methodology for engineering enzymes that can synthesize glycans. Rutgers University (RU) CBE Department Graduate Research Symposium, Apr 2019, Piscataway. (Oral Presentation by Student)
 27. **Chundawat SPS***, Nemmaru B, Hilton M, Hackl M, Lopez C, Gnanakaran S, Lang M. *Single-molecule characterization of protein adsorption to multivalent glucan polymers like cellulose*. AICHE Annual Meeting 2018, Pittsburgh. (Oral Presentation by Chundawat)
 28. Bandi CK, Goncalves A, **Chundawat SPS***. *Engineering a multifunctional family 5 glycosyl hydrolase into a transglycosidase*. AICHE Annual Meeting 2018, Pittsburgh. (Oral Presentation by Chundawat)
 29. Bandi CK*, Goncalves A, **Chundawat SPS**. *Engineering a multifunctional family 5 glycosyl hydrolase into a transglycosidase*. ACS Annual Meeting 2018, Boston. (Oral Presentation by Student)
 30. **Chundawat SPS***, Nemmaru B, Hilton M, Hackl M, Lopez C, Gnanakaran S, Lang M. *Single-molecule characterization of protein adsorption to multivalent glucan polymers like cellulose*. ACS Annual Meeting 2018, Boston. (Oral Presentation by Chundawat)
 31. **Chundawat SPS***, Sousa L, Pingali SV, O'Neill H. *In-situ small-angle neutron scattering investigation of cellulose dissolution*. ACS Annual Meeting 2018, Boston. (Oral Presentation by Chundawat)
 32. Nemmaru B, Brady SK, Lang MJ, **Chundawat SPS***. *Single-Molecule Characterization of Protein Adsorption to Multivalent Glycan Polymers*. AICHE Annual Meeting 2017, Minneapolis. (Oral Presentation by Chundawat)
 33. Farino C, Nemmaru B, **Chundawat SPS***. *Site-Directed Mutagenesis of Family 64 CBM Provides Insights into the Anomalous Binding Interactions with Pretreated Cellulose during Biomass Saccharification*. AICHE Annual Meeting 2017, Minneapolis. (Oral Presentation by Chundawat)
 34. **Chundawat SPS***, Brady S, Narayanan V, Lang M. *Providing a Molecular Basis to Modeling Protein Adsorption to Multimodal Carbohydrate-based Ligands*. AICHE Annual Meeting 2016, San Francisco. (Oral Presentation by Chundawat)
 35. Brady S*, **Chundawat SPS**, Lang M. *Cellobiohydrolase behavior visualized at the single molecule level*. Society of Industrial Microbiology and Biotechnology Annual Meeting session on "Biocatalysis: Lignocellulosic Conversion Sponsored by Novozymes" 2016, New Orleans. (Oral Presentation by Collaborator)
 36. Brady S*, **Chundawat SPS**, Lang M. *A Multi-Scale Investigation of Cellulases and Their Interaction with Cellulose Allomorphs*. AICHE Annual Meeting 2015, Salt Lake City. (Oral Presentation by Collaborator)
 37. Gnanakaran S, **Chundawat SPS***, López C, Sousa L, Dale BE, Fox BG. *Engineering More Effective Catalysts for Unnatural Allomorphs of Cellulose*. AICHE Annual Meeting 2014, Atlanta. (Oral Presentation by Chundawat)
 38. **Chundawat SPS***, Gao D, Sethi A, Crews S, Sousa L, Uppugundla N, Balan V, Gnanakaran S, Dale BE. *Increased enzyme binding to substrates does not always increase catalytic activity*. AICHE Annual Meeting 2012, Pittsburgh. (Oral Presentation by Chundawat)
 39. Dale BE*, Sousa L, **Chundawat SPS**, Balan V, Bals B. *A New Low Cost Design for the AFEX Process: Application in Distributed Biofuel Production Systems*. Symposium on Biotechnology for Fuels and Chemicals 2012, New Orleans. (Oral Presentation by PhD Advisor)
 40. Pattathil S*, **Chundawat SPS**, DeMartini JD, Li H, Miller J, Brown V, Kandemkavil S, Biswal A, Saffold T, O'Neill M, York WS, Wyman C, Mohnen D, Dixon RA, Chen F, Dale BE, Hahn MG. *Comparative*

Glycomics of Plant Biomass and Insights into Cell Wall Components that Affect Recalcitrance.

Symposium on Biotechnology for Fuels and Chemicals 2012, New Orleans. (Oral Presentation by Collaborator)

41. Sousa L*, Jin M, Uppugundla N, Bokade V, Humpula J, Gunawan C, Foston M, Azarpira A, Ralph J, Bals B, Teymouri F, **Chundawat SPS**, Dale BE, Balan V. *Extractive-AFEX (E-AFEX) Pretreatment: A unified approach for resolving bottlenecks to efficient cellulosic bioethanol production*. Symposium on Biotechnology for Fuels and Chemicals 2012, New Orleans. (Oral Presentation by Collaborator)
42. **Chundawat SPS***, Uppugundla N, Lipton M, Magyar M, Gao D, Balan V, Dale BE. *Role of enzyme blend composition, biomass particle size and solids loading on pretreated biomass saccharification efficiency*. AIChE Annual Meeting 2011, Minneapolis. (Oral Presentation by Chundawat)
43. **Chundawat SPS**, Dale BE*. *Fundamental insights to the AFEX process that catalyzes the rapid deconstruction of lignocellulose to fuels and chemicals*. 1st Brazilian BioEnergy Science and Technology Conference 2011, São Paulo (Brazil). (Oral Presentation by PhD Advisor)
44. **Chundawat SPS***, Cheh A, Sousa L, Uppugundla N, Gao D, Langan P, Gnanakaran S, Bellesia G, Agarwal U, Bianchetti C, Phillips G, Balan V, Dale BE. *Cellulose hydrolysis kinetics is closely related to its crystalline structure*. Symposium on Biotechnology for Fuels and Chemicals 2010, Clearwater. (Oral Presentation by Chundawat)
45. **Chundawat SPS***, Sousa L, Hodge D, Balan V, Dale BE. *Alkaline Based Pretreatments at Great Lakes Bioenergy Research Center*. AIChE Annual Meeting 2010, Salt Lake City. (Oral Presentation by Chundawat)
46. **Chundawat SPS***, Donohoe B, Sharma L, Elder T, Askeland P, Vismeh R, Agarwal U, Humpula J, Garlock R, Jones D, Chambliss K, Himmel M, Balan V, Dale BE. *Ultra-structural and physicochemical modifications within ammonia pretreated lignocellulosic cell walls that influence enzyme accessibility*. Symposium on Biotechnology for Fuels and Chemicals 2009, San Francisco. (Oral Presentation by Chundawat)
47. **Chundawat SPS***, Gao D, Krishnan C, Poland J, Stege J, Lipton M, Balan V, Dale BE. *Enzymatic hydrolysis of AFEX treated corn stover by cellulolytic and hemicellulolytic synergistic enzyme cocktails*. AIChE Annual Meeting 2008, Philadelphia. (Oral Presentation by Chundawat)
48. **Chundawat SPS***, Balan V, Dale BE. *Synergistic Hydrolysis of AFEX treated Corn Stover and Poplar using an Automated 96-well BCRL Microplate Assay*. AIChE Annual Meeting 2007, Salt Lake City. (Oral Presentation by Chundawat)
49. **Chundawat SPS***, Balan V, Jones D, Sousa L, Lau M, Dale BE. *Towards a Fundamental Understanding of Ammonia Fiber Expansion (AFEX) Pretreatment and its Effect on Enzymatic Hydrolysis and Fermentation*. Symposium on Biotechnology for Fuels and Chemicals 2007, Denver. (Oral Presentation by Chundawat)
50. **Chundawat SPS***, Venkatesh B, Dale BE. *Enzyme Synergies in the Hydrolysis of AFEX Pretreated Biomass*, AIChE Annual Meeting 2005, Cincinnati. (Oral Presentation by Chundawat)

Posters (asterisk* indicates poster presenter)

1. Shimabukuro S*, Garcia R, Umashankar M, DeChellis A, **Chundawat SPS**. *Catalytic Activity of Cellulolytic Enzymes, from Terrestrial Sources, on Marine Sargassum Seaweed*. Society for Industrial Microbiology and Biotechnology (SIMB) Symposium on Biomaterials, Fuels and Chemicals (SBFC) Annual Meeting 2025, May 2025, New Orleans. (Poster Presentation by Student)

2. Garcia R*, Shimabukuro S, Shen M, Dalvi N, Umashankar M, Seamann A, DeChellis A, Khare S, **Chundawat SPS**. *Bioprospecting and Characterization of Novel Marine CAZymes for the Development of a Minimal Sargassum Deconstruction Cocktail*. Society for Industrial Microbiology and Biotechnology (SIMB) Symposium on Biomaterials, Fuels and Chemicals (SBFC) Annual Meeting 2025, May 2025, New Orleans. (Poster Presentation by Student)
3. Seamann A*, Tallavajhula S, Lee A, DeChellis A, Murphy R, Garcia R, **Chundawat SPS**, Khare S. *Computational Protein Design for SaBRe*. Sargassum Biorefinery (SaBRe) Center Annual Meeting, Apr 2026, Piscataway. (Poster Presentation by Student)
4. Garcia R*, Shimabukuro S, Shen M, Dalvi N, Umashankar M, Seamann A, DeChellis A, Khare S, **Chundawat SPS**. *Characterization of Sargassum Deconstruction Enzymes and Development of Minimal Cocktail*. Sargassum Biorefinery (SaBRe) Center Annual Meeting, Apr 2026, Piscataway. (Poster Presentation by Student)
5. Umashankar M*, Garcia R, Dalvi N, **Chundawat SPS**. *Thermochemical Pretreatments Enable Rapid Sargassum Deconstruction*. Sargassum Biorefinery (SaBRe) Center Annual Meeting, Apr 2026, Piscataway. (Poster Presentation by Student)
6. Murphy R*, Seamann A, Shimabukuro S, DeChellis A, Khare S, **Chundawat SPS**. *Deep Learning Guided Design and Characterization of Sargassum Carbohydrate Binders*. Sargassum Biorefinery (SaBRe) Center Annual Meeting, Apr 2026, Piscataway. (Poster Presentation by Student)
7. Shimabukuro S*, Garcia R, DeChellis A, **Chundawat SPS**. *Catalytic Activity of Cellulolytic Enzymes, from Terrestrial Sources, on Marine Sargassum Seaweed*. Sargassum Biorefinery (SaBRe) Center Annual Meeting, Apr 2026, Piscataway. (Poster Presentation by Student)
8. Tallavajhula S*, Seamann A, Chandu A, Muthusamy S, Khare S, **Chundawat SPS**. *ProteinMPNN guided engineering of a dual functional GH29 α -L-Fucosidase from Thermotoga maritima for Improved Expression, Stability, and Activity*. Sargassum Biorefinery (SaBRe) Center Annual Meeting, Apr 2026, Piscataway. (Poster Presentation by Student)
9. DeChellis A*, Nemmaru B, **Chundawat SPS**. *Supercharging Cellulolytic Enzyme Surfaces Increases Thermostability and Catalytic Activity on Cellulosic Biomass*. AIChE Annual Meeting 2025, Boston. (Poster Presented by Student In-Person).
10. Garcia R*, Gallimore-Rapole E, Avalos J, **Chundawat SPS**. *Characterization of Sargassum Deconstruction Enzymes Using in vitro and vivo strategies*. Virtual Institute of Feedstocks of the Future (VIFF) Annual Convening Meeting, Sept 2025, Mountain View (CA). (Poster Presentation by Garcia and Gallimore-Rapole)
11. Narvekar A*, Suaini NB, Thakur R, Boulton D, Whiteley B, Giuffre B, Long W, Heacock W, Robinson R, Colangelo C, **Chundawat SPS**. *Glycosylated monoclonal antibody critical quality attributes determination using fully automated sample preparation and analytical workflows*. Bioprocess International Annual Meeting 2025, Boston. (Poster Presentation by Narvekar).
12. Salle E*, Tallavajhula S, **Chundawat SPS**. *Engineering E. coli for the Production of 2'-Fucosyllactose from Recycled Whey Lactose*. Rutgers University (RU) CBE Department Undergraduate Research Symposium, April 2026, Piscataway. (Poster Presentation by Student)
13. Padmanabhan A*, Thakur R, **Chundawat SPS**. *Quantifying the Relationship Between Optical Density and Cell Count in a Perfusion Bioreactor*. Rutgers University (RU) CBE Department Undergraduate Research Symposium, April 2026, Piscataway. (Poster Presentation by Student)

14. Semira P*, Narvekar A, **Chundawat SPS**. *Extraction and Characterization of Exosomes Isolated From Chinese Hamster Ovary Cells*. Rutgers University (RU) CBE Department Undergraduate Research Symposium, April 2026, Piscataway. (Poster Presentation by Student)
15. Garcia R*, Murphy R, Seamann A, Xie L, DeChellis A, Shimabukuro S, Khare S, **Chundawat SPS**. *Design and Discovery of Sargassum Seaweed Degrading Carbohydrate-Active Enzymes*. Gordon Research Conference (GRC) on Carbohydrate-Active Enzymes for Glycan Conversions, July 2025, Andover (NH). (Poster Presentation)
16. Murphy R*, Santo K, Hackl M, Neimark A, **Chundawat SPS**. *Unfolding & Unbinding Mechanisms of a Family 3a Carbohydrate Binding Module (CBM) from a Crystalline Cellulose Surface*. Gordon Research Conference (GRC) on Carbohydrate-Active Enzymes for Glycan Conversions, July 2025, Andover (NH). (Poster Presentation)
17. Shimabukuro S*, DeChellis A, **Chundawat SPS**. *Enzyme Engineering and Characterization for Waste Polymer Deconstruction*. Rutgers University (RU) SOE Undergraduate James J Slade Research Symposium, Apr 2025, Piscataway. (Poster Presentation by Student)
18. Seamann A*, Garcia R, Xie L, DeChellis A, Tallavajhula S, **Chundawat SPS**, Khare S. *Stabilizing Enzymes for Sargassum Biorefining*. Sargassum Biorefinery (SaBRe) Center Annual Meeting, Jan 2025, Princeton. (Poster Presentation by Student)
19. Garcia R*, Murphy R, Seamann A, Xie L, DeChellis A, Khare S, **Chundawat SPS**. *Design and Discovery of Sargassum Polysaccharides Degrading Enzymes*. Sargassum Biorefinery (SaBRe) Center Annual Meeting, Jan 2025, Princeton. (Poster Presentation by Student)
20. Narvekar A, Potter O, Heacock W, Williams S, Reddy J, Ierapetritou M, **Chundawat SPS***. *An Integrated Platform for Fully Automated, Continuous, & Real-Time Multi-Attributes Monitoring of Upstream Processes for Glycosylated Monoclonal Antibodies Production*, National Institute for Innovation in Manufacturing Biopharmaceuticals (NIIMBL) Annual Meeting 2024, Washington DC. (Poster Presentation)
21. Johnson M*, DeChellis A, Nemmaru B, **Chundawat SPS**, Lang MJ. *Processive degradation of cellulose by bacterial cellulase TfCel6B*, Biophysical Society Annual Meeting 2024, Philadelphia (PA). (Poster Presentation)
22. DeChellis A*, Murphy R, Hackl M, Ganesan G, **Chundawat SPS**. *Characterizing Multivalent Carbohydrate Binding Modules Interactions at Polysaccharide Interfaces with Acoustic Force Spectroscopy*, Biophysical Society Annual Meeting 2024, Philadelphia (PA). (Poster Presentation)
23. Murphy R*, Hackl M, Lopez C, Kolattukudy SP, Gnanakaran S, Neimark A, **Chundawat SPS**. *Acoustic Force Spectroscopy Reveals Subtle Differences in Interfacial Protein-Polysaccharide Binding Interactions*, Biophysical Society Annual Meeting 2024, Philadelphia (PA). (Poster Presentation)
24. Kumar M*, Tallavajhula SV, Burgin T, **Chundawat SPS**, *Advancing glycosynthase engineering for oligosaccharide synthesis with azido sugars leveraging transition state stabilization*, Biophysical Society Annual Meeting 2024, Philadelphia (PA). (Poster Presentation)
25. Huh H*, Lam E, **Chundawat SPS**, Lee SH, *Observing the Dynamic Process of Cellulose Polysaccharide Biosynthesis and Assembly into Cell Walls Through Live Imaging of Plant Cells*, Biophysical Society Annual Meeting 2024, Philadelphia (PA). (Poster Presentation)
26. Kumar M*, Tallavajhula SV, Burgin T, **Chundawat SPS**, *Directed evolution of glycosynthase enzymes for oligosaccharides synthesis via rapid in-vivo activity screening*, Gordon Research Conference (GRC) on Carbohydrate-Active Enzymes for Glycan Conversions, July 2023, Andover (NH). (Poster Presentation)

27. Kumar M*, Tallavajhula SV, Burgin T, **Chundawat SPS**, *Directed evolution of glycosynthase enzymes for oligosaccharides synthesis via rapid in-vivo activity screening*, Gordon Research Conference (GRC) on Carbohydrate-Active Enzymes for Glycan Conversions, July 2023, Andover (NH). (Poster Presentation)
28. DeChellis A*, Nemmaru B, Sammond D, **Chundawat SPS**, *Supercharging cellulolytic enzyme surfaces can increase thermostability & catalytic activity on cellulose and lignocellulosic biomass*, Gordon Research Conference (GRC) on Carbohydrate-Active Enzymes for Glycan Conversions, July 2023, Andover (NH). (Poster Presentation)
29. Jayachandran D*, Huh H, Lam E, Lee SH, **Chundawat SPS**, *Real-time in vivo visualization of cellulose biosynthesis and assembly as microfibrils into the plant protoplast cell wall matrix*, Gordon Research Conference (GRC) on Carbohydrate-Active Enzymes for Glycan Conversions, July 2023, Andover (NH). (Poster Presentation)
30. Jayachandran D*, **Chundawat SPS**, *Characterization of cellulose biopolymer synthesizing enzymes reconstituted in vitro*. Rutgers BME BESS Annual Meeting 2022, Piscataway. (Poster Presentation)
31. DeChellis A*, **Chundawat SPS**, *Enzyme Supercharging for improved waste polymer hydrolysis*. Rutgers BME BESS Annual Meeting 2022, Piscataway. (Poster Presentation)
32. Hackl M*, Lopez CA, Gnanakaran S, **Chundawat SPS**, *Characterizing the driving forces of protein binding to insoluble carbohydrates*. Rutgers MBGSO Annual Meeting 2022, Piscataway. (Poster Presentation)
33. Jayachandran D*, **Chundawat SPS**, *Characterization of cellulose biopolymer synthesizing enzymes reconstituted in vitro*. AIChE Annual Meeting 2021, Boston. (Poster Presented by Student In-Person).
34. Douglass J*, Nemmaru B, Shankar S, **Chundawat SPS**, *Studying the effect of cellulase supercharging on cellulosic biomass hydrolysis*. AIChE Annual Meeting 2021, Boston. (Poster Presented by Student In-Person)
35. Gyorgypal A*, Yang O, Chaturvedi A, Chopda V, Zhang H, Ierapetritou M, **Chundawat SPS**, *Model-based Prediction and Real-Time Process Monitoring For Glycosylated Biological Drugs Production*. NIH-FDA Glycoscience Research Day 2021, Virtual Meeting. (Online Presentation by Student in June 2021)
36. Nemmaru B*, Yarbrough JM, Johnson MM, Lang MM, **Chundawat SPS**, *Multimodal characterization and rational engineering of cellulolytic enzymes and cellulose-binding domains enables lowering non-productive enzyme binding to cellulose*. Society for Industrial Microbiology and Biotechnology (SIMB) Symposium on Biomaterials, Fuels and Chemicals (SBFC) Annual Meeting 2021. (Online Student Presentation in April 2021)
37. Irfan M*, Jayachandran D*, Huh H, Muchero W, Lang M, Lam E, Lee SH, **Chundawat SPS**, *Multimodal imaging of plant protoplasts with regenerating cell walls*. ACS Annual Meeting 2020, Philadelphia. (Poster Presentation Cancelled due to COVID-19)
38. Gyorgypal A*, Chopda V, Zhang H, **Chundawat SPS**, *Real-time N-linked glycans analysis to facilitate monoclonal antibody manufacturing*. ACS Annual Meeting 2020, Philadelphia. (Poster Presentation Cancelled due to COVID-19)
39. Nemmaru B*, Ramirez N, Kravchenko N, **Chundawat SPS**, *Reducing non-productive binding of cellulases to improve crystalline cellulose hydrolysis rates*. ACS Annual Meeting 2020, Philadelphia. (Poster Presentation Cancelled due to COVID-19)

40. Chopda V, Gyorgypal A, Zhang H, **Chundawat SPS**, Hausner D*. Integration of Process Analytics for Real-Time monitoring of Critical Quality Parameters and Attributes for Continuous Biomanufacturing. International Foundation Process Analytical Chemistry (IFPAC) Annual Meeting, Feb 2020, North Bethesda. (Poster Presentation by Staff on behalf of student)
41. Gyorgypal A*, Chopda V, Zhang H, **Chundawat SPS**. At-Line N-linked Glycan Analysis to Facilitate Monoclonal Antibody Manufacturing. Rutgers University (RU) BME Department Graduate Research Symposium, Dec 2019, Piscataway. (Poster Presentation by Student)
42. Yang O*, Chopda V, Muddu SV, Gyorgypal A, Hausner D, Singh R, Tsilomelekis G, Zhang H, **Chundawat SPS**, Ramachandran R, Ierapetritou M. *Continuous Upstream Biopharmaceuticals Manufacturing*. Advanced Process Modelling (APM) Forum 2019, New York. (Poster Presentation by Student)
43. Thokkadam A*, Wang A, Nemmaru B, **Chundawat SPS**. Studying the Effect of CBM Supercharging on Endo-Exo Cellulase Hydrolysis Activity. Rutgers University (RU) SOE Undergraduate James J Slade Research Symposium, Apr 2019, Piscataway. (Poster Presentation by Student)
44. Lee S-H, **Chundawat SPS***, Lam E, Fabris L, Lang M, Muchero W, Pingali SV. *In planta multimodal single-molecule imaging of cell wall synthesis*. Department of Energy Annual PI Meeting 2019, Vienna. (Poster Presentation by Co-PI Chundawat)
45. Ma S*, Hackl M, **Chundawat SPS**. *Developing an Acoustic Force Spectroscopy Technique to Measure Carbohydrate Binding Module Interaction with Cellulose*. Rutgers University (RU) Aresty Undergraduate Research Symposium, August 2018, Piscataway. (Poster Presentation by Student)
46. Agrawal A*, Bandi CK, **Chundawat SPS**. High-throughput cell screening methodology for the directed evolution of glycosynthases. Rutgers University (RU) Society of Women Engineers (SWE) Research Symposium, Apr 2018, Piscataway. (Poster Presentation by Student)
47. Goncalves A*, Bandi CK, **Chundawat SPS**. Understanding the Role of CBMs in the Transglycosylation Efficiency of GH5 Family Enzymes. Rutgers University (RU) SOE Undergraduate James J Slade Research Symposium, Apr 2018, Piscataway. (Poster Presentation by Student)
48. Esposito B*, **Chundawat SPS**. *Ammonia Pretreatment of Cellulosic Biomass Increases Efficiency of Biofuels Production*. Rutgers University (RU) Aresty Undergraduate Research Symposium, April 2018, Piscataway. (Poster Presentation by Student)
49. Thokkadam A*, Bandi CK, Vankayalapati NS, **Chundawat SPS**. *Impact of Cellulase Supercharging on Binding to Lignocellulosic Biomass*. AIChE Northeast Regional Undergraduate Meeting 2018, San Francisco. (Poster Presentation by Undergraduate Student)
50. Liu Y*, Nemmaru B, **Chundawat SPS**. Characterization of T. fusca cellulase activity on unnatural cellulose allomorphs. Rutgers University (RU) CBE Department Graduate Research Symposium, Nov 2017, Piscataway. (Poster Presentation by Student)
51. Gupta S*, Bandi CK, Thokkadam A, **Chundawat SPS**. Developing designer glycoligands for multimodal protein bioseparations. Rutgers University (RU) CBE Department Graduate Research Symposium, Nov 2017, Piscataway. (Poster Presentation by Student)
52. Vankayalapati N*, Bandi CK, Thokkadam A, **Chundawat SPS**. Effect of supercharging cellulases on binding to cellulose and pretreated cellulosic biomass. Rutgers University (RU) CBE Department Graduate Research Symposium, Nov 2017, Piscataway. (Poster Presentation by Student)
53. Subhash N*, Bandi CK, **Chundawat SPS**. Enzymatic Synthesis of Oligosaccharides for Modulating Growth of the Gut Microbiome. Rutgers University (RU) CBE Department Graduate Research Symposium, Nov 2017, Piscataway. (Poster Presentation by Student)

54. Agrawal A*, Bandi CK, **Chundawat SPS**. High-throughput cell screening methodology for the directed evolution of glycosynthases. Rutgers University (RU) CBE Department Graduate Research Symposium, Nov 2017, Piscataway. (Poster Presentation by Student)
55. Dagia A*, Nemmaru B, **Chundawat SPS**. Impact of a conserved tyrosine residue on Family 1 CBM binding to cellulose allomorphs. Rutgers University (RU) CBE Department Graduate Research Symposium, Nov 2017, Piscataway. (Poster Presentation by Student)
56. Samantha Cobos*, Bhargava Nemmaru, **Chundawat SPS**. *Understanding protein adsorption to TEMPO-oxidized cellulose microfibrils*. ACS Annual Meeting, April 2017, San Francisco. (Poster Presentation by Student)
57. Thokkadam A*, Bandi C, **Chundawat SPS**. *Effect of nucleophilic mutations on hydrolytic and transglycosylation activities of carbohydrate-active enzymes (CAZymes)*. Rutgers University (RU) CBE Department Undergraduate Research Symposium, April 2017, Piscataway. (Poster Presentation by Student)
58. Berger M*, Bandi C, **Chundawat SPS**. *Negatively charging cellulolytic enzymes can reduce non-productive binding to lignin*. Rutgers University (RU) Aresty Undergraduate Research Symposium, April 2017, Piscataway. (Poster Presentation by Student)
59. Park J*, McGinley E, Bandi C, **Chundawat SPS**. *Expression and Purification of CelE-CBM3a and its Mutants*. Rutgers University (RU) CBE Department Undergraduate Research Symposium, Apr 2016, Piscataway. (Poster Presentation by Student)
60. Farino C*, Nemmaru B, **Chundawat SPS**. *Binding Affinity of Family 64 Carbohydrate Binding Modules and its Mutants to Crystalline Cellulose Allomorphs*. Rutgers University (RU) Aresty Undergraduate Research Symposium, April 2017, Piscataway. (Poster Presentation by Student)
61. Bandi C*, Sahithi N, Berger M, Park J, Whitehead T, **Chundawat SPS**. *Minimizing non-productive binding of cellulases to lignin by supercharging enzyme surfaces*. Gordon Research Conference (GRC) on Cellulases and Carbohydrate-Active Enzymes, July 2017, Andover. (Poster Presentation by Student)
62. Park J*, McGinley E, **Chundawat SPS**. *Expression and Characterization of Fungal Cellulase Catalytic and Cellulose Binding Domains for Single-Molecule Assays*. Rutgers University (RU) CBE Department Undergraduate Research Symposium, Apr 2016, Piscataway. (Poster Presentation by Student)
63. Gerszberg J*, **Chundawat SPS**. *t-Test Stepping Algorithm and Variable Stepsize Hidden Markov Models for Characterizing Enzyme Motility Determined Using Optical Tweezers*. Rutgers University (RU) CBE Department Undergraduate Research Symposium, Apr 2016, Piscataway. (Poster Presentation by Student)
64. Farino C*, Mainardi D, **Chundawat SPS**. *Determining the Binding Affinity of Carbohydrate Binding Modules to Crystalline Cellulose*. Rutgers University (RU) CBE Department Undergraduate Research Symposium, Apr 2016, Piscataway. (Poster Presentation by Student)
65. Lemenze A*, **Chundawat SPS**. *Rutgers Chem-E-Car: The Drunken Snail*. Rutgers University (RU) CBE Department Undergraduate Research Symposium, Apr 2016, Piscataway. (Poster Presentation by Student)
66. Desai A*, Webb AP, Bertram R, Ghosh S, Jeong E, Srivastav A, **Chundawat SPS**. *Validation of an Integrated Lignocellulosic Biorefinery Model using Aspen*. Rutgers University (RU) CBE Department Undergraduate Research Symposium, Apr 2016, Piscataway. (Poster Presentation by Student)

67. **Chundawat SPS***. *A multiscale approach to studying conversion of lignocellulosic biomass to biofuels*. Rutgers University (RU) First Annual Sustainable Symposium, April 2016, Piscataway. (Poster Presentation by Chundawat)
68. Roy S*, Berger M, Gupta S, Sousa L, **Chundawat SPS**. *Novel Pretreatment Process Enhances Cellulosic Biofuel Production*. Rutgers University (RU) CBE Department Graduate Research Symposium, Dec 2016, Piscataway. (Poster Presentation by Postdoc)
69. Bandi C*, **Chundawat SPS**. *Engineering multifunctional glycosynthases for glycan synthesis*. Rutgers University (RU) CBE Department Graduate Research Symposium, Dec 2016, Piscataway. (Poster Presentation by Student)
70. Nemmaru B*, Doran P, Liu Y, Brady S, Lang MJ, **Chundawat SPS**. *Single-molecule characterization of processive glycosyl hydrolase enzymes*. Rutgers University (RU) CBE Department Graduate Research Symposium, Dec 2016, Piscataway. (Poster Presentation by Student)
71. Haarmeyer C, Smith M, **Chundawat SPS**, Sammond DW, Whitehead T*. *Using Protein Design to Evaluate the Relationship Between Protein Surface Potential and Protein-Lignin Binding for the Eventual of Low Lignin Binding Cellulases*. AIChE Annual Meeting 2016, San Francisco. (Poster Presentation by Collaborator)
72. Poster Presentation by **Shishir Chundawat** at Gordon Research Conference/Seminar on Cellulosomes, Cellulases & Other Carbohydrate Modifying Enzymes, *Native cellulases have abnormal interactions with unnatural cellulose allomorphs*, Andover (NH), Aug 2015.
73. Berger M*, Alves K, **Chundawat SPS**. *Novel Plant Biomass Pretreatments For Enhancing Biofuel Production*. Rutgers University (RU) Aresty Undergraduate Research Symposium, Aug 2015, Piscataway. (Poster Presentation by Student)
74. Asthana S*, **Chundawat SPS**. *Characterization of the Binding Affinity of Engineered Carbohydrate-Binding Modules to Lignocellulosic Biomass*. Rutgers University (RU) CBE Department Graduate Research Symposium, Nov 2015, Piscataway. (Poster Presentation by Student)
75. Sharma S*, **Chundawat SPS**. *Using Confocal Imaging to Study Binding Dynamics of Cellulolytic Enzymes to Cellulose*. Rutgers University (RU) CBE Department Graduate Research Symposium, Nov 2015, Piscataway. (Poster Presentation by Student)

Other Interviews, Presentations, and Public Outreach

1. Interviewed in 2026 by NIIMBL for real-time monitoring PAT tools developed by Rutgers-led team to enable advanced biologics manufacturing in 2025-2026 NIIMBL annual report.
2. Interviewed with Nautilus magazine in a special issue ([Catalysts of Discovery](#)) highlighting Chundawat's participation in Schmidt Sciences SaBRE center in Oct 2025 showcasing advances in research on *Sargassum* (<https://nautil.us/new-life-for-rotting-seaweed-1238008>). Special issue was launched and made available to the public in Nov 2025 at an event in New York City.
3. Interviewed with Rutgers highlighting Chundawat's participation in RCEI symposium in May 2025 showcasing advances in research on Caribbean seaweeds (<https://rcei.rutgers.edu/collaborative-advances-in-research-on-seaweeds-in-the-caribbean/>).
4. Interviewed with Rutgers as part of press release highlighting Chundawat's 2025 Science Advances article on visualizing cellulose biosynthesis and plant cell wall assembly for the first time (<https://www.rutgers.edu/news/scientists-witness-plant-cells-generate-cellulose-and-form-cell->

- [walls-first-time](https://www.sciencenews.org/article/live-plant-cells-build-walls-grow)). Paper was highlighted online by multiple news and media agencies (e.g., <https://www.sciencenews.org/article/live-plant-cells-build-walls-grow>).
5. Press coverage in 2024 by Schmidt Sciences, and other online media, for new [Sargassum BioRefinery or SaBRe center](#) funded at Rutgers as part of Schmidt Sciences' [Virtual Institute on Feedstocks of the Future](#) (VIFF) (<https://soe.rutgers.edu/news/transforming-troublesome-seaweed-feedstock-future>).
 6. Press coverage in 2022 by NIIMBL, and other online media, for new biologics manufacturing research grant from NIIMBL Project Call 5.2T (<https://www.niimbl.org/projects/an-integrated-platform-for-fully-automated-continuous-real-time-multi-attributes-monitoring-of-upstream-processes-for-glycosylated-monoclonal-antibodies-production/>).
 7. Interviewed with Rutgers as part of press release highlighting Chundawat's 2022 PNAS article (<https://www.rutgers.edu/news/rutgers-researchers-develop-method-single-molecule-precision-engineer-enzyme-stickiness>).
 8. Interviewed with Rutgers as part of press release highlighting Chundawat's 2022 ACS Analytical Chemistry cover article (<https://newbrunswick.rutgers.edu/news/rutgers-pair-creates-monitoring-toolkit-speed-production-biologic-drugs>).
 9. Interviewed with Drug Target Review (DTR) trade magazine in Fall 2021 highlighting how Rutgers group led by Chundawat developed a new process analytical technology (PAT) system for the automated monitoring of antibody N-glycosylation to enable biologics manufacturing. Article available online on DTR website (<https://www.drugtargetreview.com/article/99976/ensuring-the-safety-and-efficacy-of-biologics-products/>).
 10. Op-Ed article titled '*Forward to the Future: Cheap, Plant-Based Materials Coming Thanks to Bioengineering Revolution*' co-authored along with Rutgers-New Brunswick's communications team on "*Engineering Fields That Will Change the World*" essay package submitted to [NJ.com](#) and the Star-Ledger for publication in Fall 2020. Copy of article is available on CBE Website (<https://cbe.rutgers.edu/news/forward-future-cheap-plant-based-materials-coming-thanks-bioengineering-revolution>).
 11. Interviewed with journalist Todd Bates in Rutgers Today and as part of ORNL press release highlighting Chundawat's 2020 Green Chemistry article (<https://neutrons.ornl.gov/content/how-make-it-easier-turn-plant-waste-biofuels>).
 12. Interviewed with Burt Sempier of WMBCTV showcasing ongoing bioenergy research being conducted at Rutgers Chundawat Lab on Feb 6th 2020: <https://wmbctv.com/portfolio-view/biofuels/>.
 13. Fall 2019 CBE newsletter article titled '*How Times Have Changed: CBE Flashback*' co-authored with Dr. Rohit Ramachandran and SOE communications team ([Fall 2019 CBE Newsletter](#))
 14. Video interview with SOE communications team in Fall 2017 to provide promotional advertising material on relevance of chemical engineering to society and overview to CBE department at Rutgers University ([YouTube Video posted on CBE Homepage](#))
 15. Cover story about Chundawat Research Group and interview in SOE's RU Engineer magazine (Winter 2017, Vol. 2, No. 1)

16. Interview with ethanol manufacturer POET in 2017 'Vital' magazine as part of a press release online article (<http://vitalbypoet.com/stories/poet-internship-program-offers-opportunity-for-experience>) highlighting Rutgers CBE student (James Forder) internship at POET.
17. Interviewed on WRNJ Radio (at 11:15 am on July 7th 2017) by Ms. Joyce Estey (WRNJ Radio News Director) to discuss on-going research at Rutgers University on how cellulases.
18. Interviewed with science journalist Todd Bates in Rutgers Today as part of a press release online article (<http://news.rutgers.edu/research-news/cutting-cost-ethanol-other-biofuels-and-gasoline/20170630#.WZ3bXq2ZMmJ>) that highlighted a journal cover article on cellulases.
19. Press coverage in 2016 for journal cover article highlighted by The Royal Society of Chemistry (<http://cbe.rutgers.edu/news/chundawat-group-research-highlighted-royal-society-chemistry-journal-cover-article>).
20. Press coverage in 2016 for journal article highlighted by U.S. Department of Energy Bioenergy Center (<https://www.glbrc.org/research/highlights/next-generation-ammonia-pretreatment-enhances-cellulosic-biofuel-production>).
21. Press coverage in 2016 on collaborative *Nature Communications* article highlighted in social media (<http://www.labmanager.com/news/2015/12/anatomy-of-a-microscopic-wood-chipper#.VnB5LYTWpW8>).
22. Interviewed for coverage on sustainable biofuels focused on 2013 *PNAS* article highlighted in social media (<http://www.biofuelsdigest.com/bdigest/2013/07/02/researchers-find-new-method-to-convert-cellulose-for-better-biofuel-production/>).
23. Press coverage on *Energy and Environmental Science* article in 2011 by The Royal Society of Chemistry and U.S. DOE GLBRC (<http://blogs.rsc.org/ee/2011/01/11/improving-pre-treatments-for-better-biomass-conversion/>).
24. Interviewed for press coverage on *JACS* article on cellulosic biofuels highlighted in *AIChE Chemical & Engineering Progress Magazine* in September Issue (2011, Vol. 107, No. 9) and other social media (<http://greatlakesecho.org/2011/10/26/research-may-strengthen-biofuel-production-from-non-edible-sources/>).
25. Interviewed for press coverage on *J. Phys. Chem. A* journal cover article in 2011 (<http://pubs.acs.org/doi/abs/10.1021/jp203620x>).

Service Activities

Service to Rutgers University

2025-2026: Co-Chair, CBE Faculty Search Committee

2024-ongoing: Member, First Year CBE/SOE IDEAS Lab Core Committee to Increase Engagement

2023-2024: Faculty Committee Member, Provost's Strategic Task Force on the Life Sciences Alliance

2023-11: Lead Faculty, SOE Freshman Orientation lecture series representing CBE Department. Developed orientation presentation and hands-on demo activity for 8 x 80 min lectures.

2023-2025: Supported recruitment of visiting faculty (Anurag Rathore) at CBE Department and launch of a new summer course focused on biopharmaceuticals manufacturing starting in 2023.

2023-ongoing: Co-Organizer (with Dr. Gregory Dignon), CBE Annual Fall/Spring Seminar Series

2020-ongoing: Member, SLAM-Rutgers Strategic Committee for Design of New EP2/CBE building

2015-ongoing: CBE Department Website & Social Media Activities (e.g., Twitter) Committee

2021-2024: Member, CBE Graduate Student Admissions Ad Hoc Review Committee

2021-2022: Chair, CBE Faculty Search Committee

2021-08: Chair, CBE Biochemical Engineering Working Group at Annual CBE Faculty Retreat

2021-03: Member, CBE Representative for Rutgers Giving Day Fund Raising Event for EP2 Building

2020-12: Chair, CBE Department Microsoft Teams Introduction and CBE Channel Launch

2020-2023: Co-Organizer (with Dr. Benjamin Schuster), CBE Annual Fall/Spring Seminar Series

2015-2019: Undergraduate Faculty Advisor for American Institute of Chemical Engineers (AIChE)
Rutgers Student Chapter Chem-E-Car Team

2019: Coordinated donations and no-cost loans to Rutgers, together with Dr. Doug Hausner and Dr. Haoran Zhang, of equipment and supplies worth approximately \$450,000 from Agilent, Eppendorf, and Genscript to establish first-of-its-kind biologics manufacturing R&D facility at Rutgers University.

2019-2020: Member, CBE Faculty Search Committee

2019: Invited Ad Hoc Reviewer for Rutgers Vice Chancellor for Research and Innovation Office

2019: Member, CBE-Rutgers Strategic Planning Sub-Committee on CBE/SOE Infrastructure

2018: CBE Committee Member, Graduate Certificate/Concentration in Biochemical Engineering

2017: Speaker, Invited faculty panel speaker at Rutgers Global (GAIA) Grant Symposium on Nov 16th

2017-2018: Chair, Masters Certificate/Concentration in Energy CBE Committee

2017-2018: Chair, CBE Faculty Search Committee

2017: Faculty representative for outreach and support to Chem-E-Car team at AIChE Regional Meeting

2017: Faculty panelist on Engineering & Medicine for admitted Rutgers students

2017: Faculty representative for outreach and representation at NJ Future City Competition

2017: Education and outreach for local NJ high-school students on July 14th, 2017

2017: Faculty representative for CBE outreach, Rutgers Day 2017

2017: Member, SOE Freshman Orientation lecture series representing CBE Department

2017: Organizing tour of research labs for admitted undergraduate students and families

2016: Coordinated donations to Rutgers of equipment and supplies worth approximately \$250,000 from GE Healthcare (Piscataway, NJ).

2016-2017: Member, CBE Faculty Search Committee

2016: Organized a half-day seminar introduction to Chemical Engineering and conducted a hands-on Chem-E-Car design activity for high school students and parents on July 27th, 2016

2016: Organized a half-day seminar introduction to Chemical Engineering and conducted a hands-on Chem-E-Car design activity for pre-engineering summer academy held on Aug 5th, 2016

2016: Faculty representative for CBE outreach, Rutgers Day 2016

2016: Member, CBE Qualifying Exam Committee

2016: Member, CBE-Rutgers Strategic Planning Sub-Committee on Department Infrastructure

2015-2018: Faculty Advisor, CBE Undergraduate Junior Students

2015-2019: Member, CBE Graduate Student Admissions Review Committee

2015-2019: Member, CBE Undergraduate Curriculum Committee

2015-2019: Faculty Advisor, AIChE Rutgers Chapter ChemE Car Team

2015: Judge, CBE Graduate Research Symposium

2015: Member, Masters of Engineering (ME) in Energy Systems CBE Sub-Committee

2015: Member, CBE Qualifying Exam Committee

Other Academic Service at Rutgers

1. Invited CBE department speaker at Rutgers annual SOE Women in Engineering event for ~30 High School Female Students (March 1st, 2019)
2. Invited speaker and guest panelist at Rutgers annual CABLE (Consortium for Advanced Bioeconomy Leadership Education) event on 'Transforming Transportation for a Low-Carbon Future' hosted by Dr. Serpil Guran at Livingston Student Center for ~20 Rutgers Undergraduates (March 7th, 2019)
3. AIChE ChemE Car team faculty co-advisor with Dr. Alex Bertuccio, 2019, 15 advisees per year
4. Introduction to CBE seminar followed by hands-on cellulosic biofuels process design activity for ~45 High School Students as part of 2018 half-day (July 20th) pre-engineering summer academy sponsored by Rutgers University (Division of Continuing Studies)
5. Basic introduction to Foldit online protein modeling/gaming software and 90 mins hands-on training session for volunteer CBE undergraduate students at Rutgers CBE (March 20th 2018)
6. AIChE ChemE Car team faculty advisor, 2018, 15 advisees per year
7. 2017 Introduction to CBE seminar followed by hands-on cellulosic biofuels process design activity for ~45 High School Students as part of 2017 half-day (July 14th) pre-engineering summer academy sponsored by Rutgers University (Division of Continuing Studies)
8. AIChE ChemE Car team faculty advisor, 2017, 15 advisees per year
9. CBE faculty advisor for junior year students, 2017, 119 advisees
10. Introduction to CBE seminar followed by hands-on CBE Rocket-Car design activity, and tour of CBE department for ~50 High School Students and their parents as part of 2016 half-day (July 27th) introduction to engineering summer outreach program sponsored by Rutgers School of Engineering
11. Introduction to CBE seminar followed by hands-on CBE Rocket-Car design activity for ~40 High School Students as part of 2016 half-day (Aug 5th) pre-engineering summer academy sponsored by Rutgers University (Division of Continuing Studies)
12. AIChE ChemE Car team faculty advisor, 2016, 15 advisees per year
13. CBE faculty advisor for junior year students, 2016, 109 advisees

14. AIChE ChemE Car team faculty advisor, 2015, 15 advisees per year

Service to Other Public Bodies

1. Invited guest panelist at Great Lakes Bioenergy Research Center's (GLBRC's) annual networking event with alumni hosted by Dr. Yiying Xiong with several GLBRC staff (March 18th, 2025)
2. 2025: National Science Foundation (NSF) Research Capacity and Competitiveness Section (OD/OIA/RCC) Program Invited Panel Reviewer
3. 2024: Sun Grant Program (SGP), National Institute of Food and Agriculture (NIFA), U.S. Department of Agriculture (USDA), Invited Ad Hoc Panel Reviewer
4. 2024: National Science Foundation (NSF) Chemical, Bioengineering, Environmental, and Transport Systems (CBET) Program Invited Ad Hoc Panel Reviewer
5. 2023: National Science Foundation (NSF) Molecular and Cellular Biosciences (MCB) Program Invited Panel Reviewer
6. 2023: National Science Foundation (NSF) Chemical, Bioengineering, Environmental, and Transport Systems (CBET) Program Invited Panel Reviewer
7. 2023: National Science Foundation (NSF) Chemical, Bioengineering, Environmental, and Transport Systems (CBET) Program Invited Ad Hoc Panel Reviewer
8. 2023: National Science Foundation (NSF) Chemistry (CHE) Program Invited Ad Hoc Panel Reviewer
9. 2022: National Science Foundation (NSF) Chemical, Bioengineering, Environmental, and Transport Systems (CBET) Program Invited Ad Hoc Panel Reviewer
10. 2021: National Science Foundation (NSF) Chemical, Bioengineering, Environmental, and Transport Systems (CBET) Program Invited Panel Reviewer
11. 2020: U.S. Civilian Research and Development Foundation for the Independent States of the Former Soviet Union (CRDF) Global Grant Program Invited Panel Reviewer
12. 2020: National Science Foundation (NSF) Chemical, Bioengineering, Environmental, and Transport Systems (CBET) Program Invited Panel Reviewer
13. 2018: Swedish Research Council (Vetenskapsrådet) Invited Proposal Reviewer
14. 2018: National Science Foundation (NSF) Chemical, Bioengineering, Environmental, and Transport Systems (CBET) Program Invited Panel Reviewer
15. 2015: National Science Foundation (NSF) Chemical, Bioengineering, Environmental, and Transport Systems (CBET) Program Invited Panel Reviewer
16. 2013: Invited Reviewer for Department of Energy (DOE) Small Business Innovation Research and Small Business Technology Transfer Program (SBIR/STTR)
17. 2013: Invited Reviewer for Doctoral Fellowship Program of the Austrian Academy of Sciences
18. 2010: European Research Area Industrial Biotechnology (ERA-IB) Program Invited Proposal Reviewer

Rutgers Graduate Program Affiliations

1. Rutgers Graduate School - New Brunswick, Full Member
2. Rutgers Climate and Energy Institute - New Brunswick, Faculty Member
3. Rutgers Graduate Program in Quantitative Biomedicine - New Brunswick, Full Member
4. Rutgers Graduate Programs in Molecular Biosciences - New Brunswick, Full Member
5. Rutgers Graduate Program in Biomedical Engineering - New Brunswick, Associate Member

6. Rutgers Energy Institute - New Brunswick, Faculty Member
7. Rutgers Center for Nutrition, Microbiome, and Health - New Brunswick, Faculty Member
8. Rutgers Graduate Program in Chemical and Biochemical Engineering - New Brunswick, Full Member

Other Professional Activities

Participation in Organizing or Chairing Conferences, Workshops, and Organizations

1. 2026: Sargassum BioRefinery (SaBRe) Annual Meeting Organizing Chair at Rutgers University sponsored by Schmidt Sciences.
2. 2011-2022: Chair/Co-chair of the '*Biological Conversions and Processes for Renewable Feedstocks*' session at American Institute of Chemical Engineers (AIChE) Annual Meeting sponsored by the Sustainable Engineering Forum.
3. 2020: Invited virtual panel participant on "*Pharmaceuticals And BioTechnology: Biotherapeutics and Biosimilars (Session V15H1S2)*" moderated by Prof. Anurag Rathore and sponsored by Vaishwik Bharatiya Vaigyanik or VAIBHAV Virtual Summit (Government of India)
4. 2016: Invited panelist/participant on "*Papertronics: Paper-based Electronics for the 21st Century*" hosted by Prof. Aaron Mazzeo and sponsored by NSF in Arlington (VA)
5. 2011: Retreat Planning Committee Area 2 Lead at Department of Energy (DOE) sponsored Great Lakes Bioenergy Research Center Annual Retreat in South Bend (Indiana)

Academic Journals Peer-Review and Editorial Board Service

List of Journals Served to-date as Invited Reviewer (since 2005)

ACS Biochemistry, ACS Catalysis, ACS Nano, ACS Omega, ACS Sustainable Chemistry & Engineering, ACS Synthetic Biology, AIChE Journal, Applied Biochemistry and Biotechnology, Biochemical Engineering Journal, Biomacromolecules, Biomass and Bioenergy, Bioresource Technology, BioResources, Biotechnology Advances, Biotechnology and Applied Biochemistry, Biotechnology for Biofuels, Biotechnology Progress, Carbohydrate Polymers, Cellulose, Chemical and Biochemical Engineering Quarterly, Chemistry – A European Journal, Chemical Reviews, Energy and Environmental Science, Energy and Fuels, Energy, Enzyme and Microbial Technology, Frontiers, Journal of Chemical Technology & Biotechnology, Microbial Cell Factories, Nature Materials, PLoS ONE, Proceedings of the National Academy of Sciences USA, Process Biochemistry, RSC Green Chemistry, Science Advances, The Plant Journal, Wiley Biotechnology and Bioengineering, Food Bioscience

Associate editor (since 2021)

Associate editor for *Frontiers in Molecular Biosciences* journal (Specialty Section: Glycoscience). See journal weblink (<https://www.frontiersin.org/journals/molecular-biosciences/sections/glycoscience>).

Review editor (since 2011)

Served as invited review editor for *Frontiers in Energy Research* and *Frontiers in Bioengineering and Biotechnology* journals (Specialty Section: Bioenergy & Biofuels). See journal weblink for responsibilities of review editor (<http://home.frontiersin.org/about/publishing-model>).

List of Open Access Refereed Articles for Frontiers in Energy Research since 2015:

- <https://doi.org/10.3389/fimmu.2024.1347871>
- <https://doi.org/10.3389/fenrg.2021.680313>
- <https://doi.org/10.3389/fenrg.2017.00005>
- <https://doi.org/10.3389/fmars.2017.00293>
- <https://doi.org/10.3389/fenrg.2018.00067>

Professional Societies Member

Member of American Chemical Society (ACS)

Member of American Institute of Chemical Engineers (AIChE)

Member of Biophysical Society (BPS)